Understanding the Basics of Environmental Monitoring

David Matsuhiro Cleanroom Compliance Inc. dmatsuhiro@cleanroomcompliance.com

- Surfaces types include walls, floors, doors, windows, equipment, and tanks
- Types of Test Methods
 - RODAC, Replicate Organism Detection And Counting Plates
 - Flexible Films
 - Swabs
 - Surface Rinse Methods

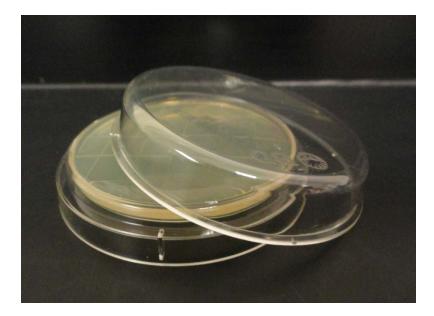
RODAC Plates

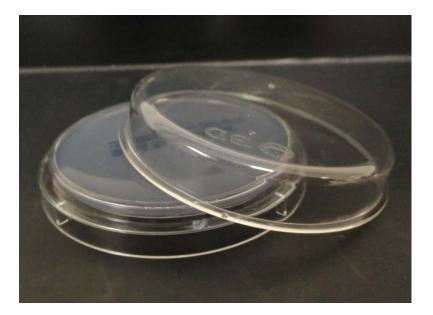
- A 50 mm plate is filled with media so that the agar extends above the plate
- The surface of the media is rolled against a flat surface to sample
- The contact surface area of a RODAC is 25 cm²
- This is the most common method for sampling surfaces

RODAC Plates

- > Automated sampling devices are available
- Neutralizers are usually within the media
- Available, single, double or triple bagged and gamma irradiated
- Recommend not using plates poured in a Grade A area, then bagged aseptically

RODAC Plates

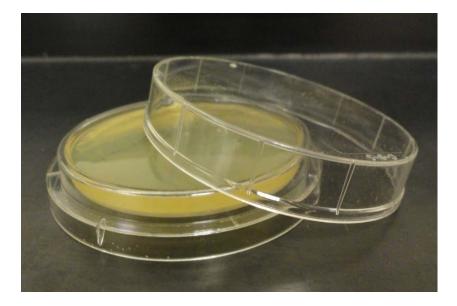


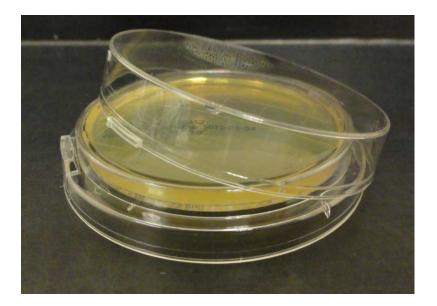


Poly-80 & Lec

D/E Neutralizer

RODAC Plates

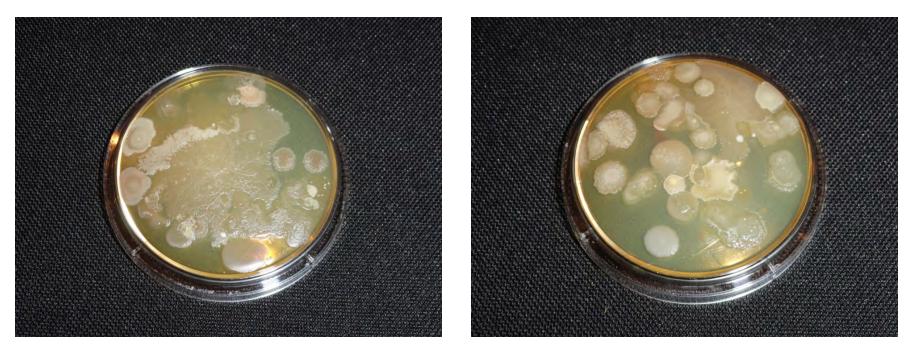




Tacky Plates

Locking Plates

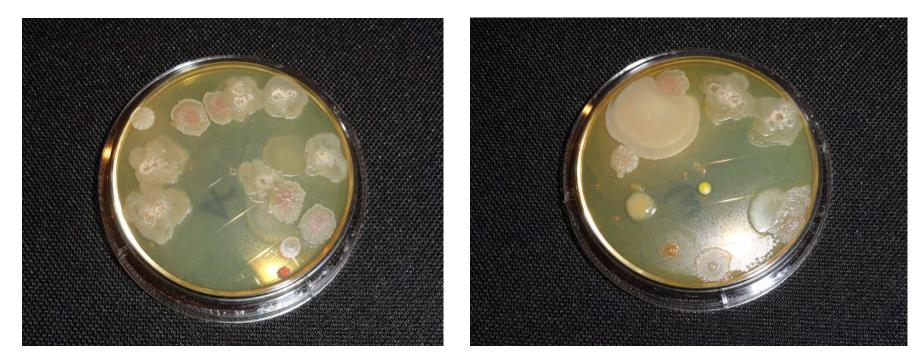
RODAC Plates Results



First Sample

Second Sample

RODAC Plates Results



Third Sample

Fourth Sample

RODAC Plates Results



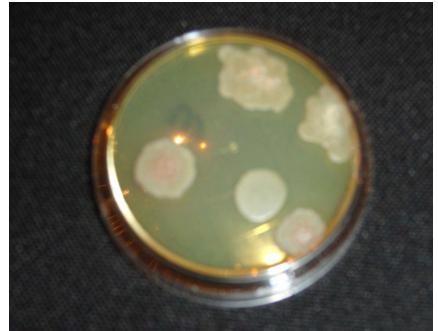


Sixth Sample

Fifth Sample

RODAC Plates Results





Seventh Sample 10

Eighth Sample

RODAC Contact Plates

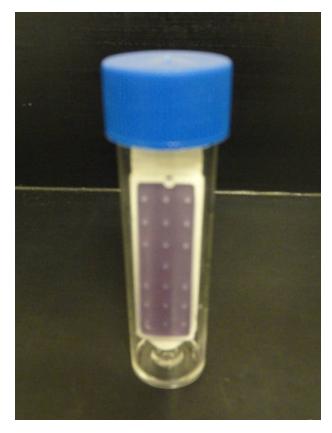
- Advantages
 - Ease of use
 - Fast
 - Economical

- Disadvantages
 - Not suitable for irregular surfaces
 - Media residue must be removed from the sample site
 - Recovery rate very low

Flexible Film

- Media is deposited on a flexible material and used like a contact plate
- Can be used on irregular surfaces
- There are several versions, most have a surface area of 25 cm²

Flexible Film





Flexible Film





Flexible Film Contact Strips

- Advantages
 - Ease of use
 - Economical
 - Can bend in several configurations

- Disadvantages
 - Not suitable for irregular surfaces
 - Media residue must be removed after sampling
 - Recovery rate very low

Swab Testing

- > Used for sampling irregular surfaces
- A sterile dry/moist swab is rubbed over 25 cm², if possible
- Swabs may be made of Cotton, Dacron, Calcium Alginate or acceptable material

Swab Testing

- > Organisms are transferred from the surface tested to the swab or paddle
 - Liquid Method
 - The swab is broken off into 3 5 ml of neutralizing broth
 - The broth/swab is vortexed to remove the organisms from the swab

Liquid Method Continued

- The organisms are transferred from the swab to the neutralizing broth
- The broth is plated leaving the swab in tube

Direct to Broth Swab Method

- After sampling the swab is broken directly into TSB
- The swab is directly inoculated into the media
 - This method is qualitative and not quantative
 - Fast and easy to use

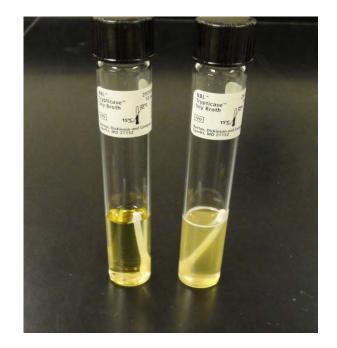
Direct to Broth Swab Method





Direct to Broth Swab Method





Direct to Agar Swab Method
The swab is directly inoculated on a TSA plate containing a neutralizer
This method is quantative
Fast and easy to use

Direct to Agar Method

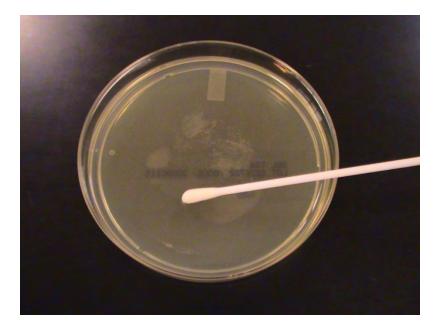






Direct to Agar Method





Direct Agar Method





First Sample

Second Sample

Direct Agar Method

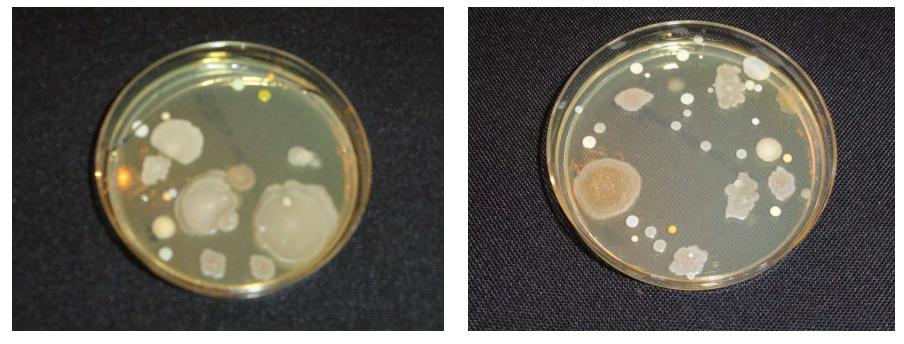




Third Sample

Forth Sample

Direct Agar Method



Fifth Sample

Sixth Sample

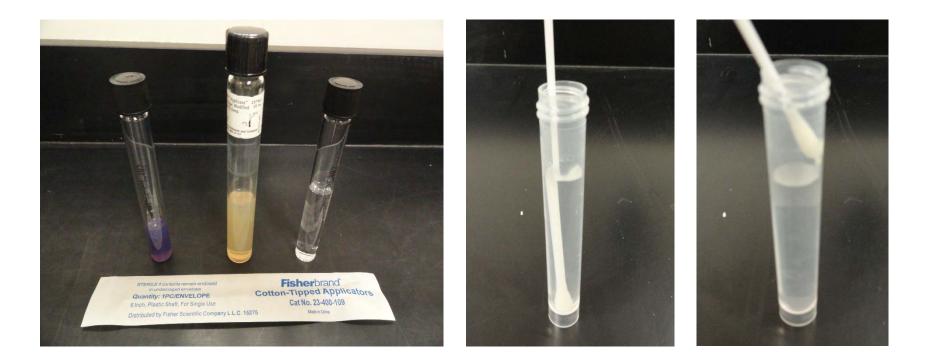
Direct Agar Method



Seventh Sample Note Spreading Bacillus

Pour Plate Method

- Swab is wetted
- Excess liquid is squeezed out
- Surface is sampled
- Swab is placed into a neutralizing broth
- The broth is mixed and poured into an Perti Dish
- Molten agar is poured into the plate and mixed



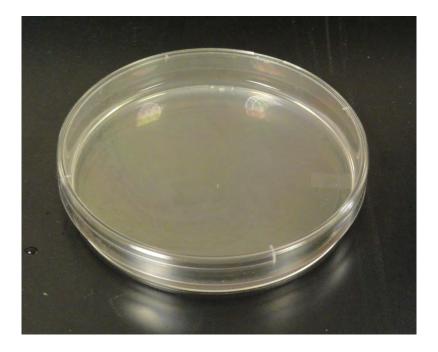




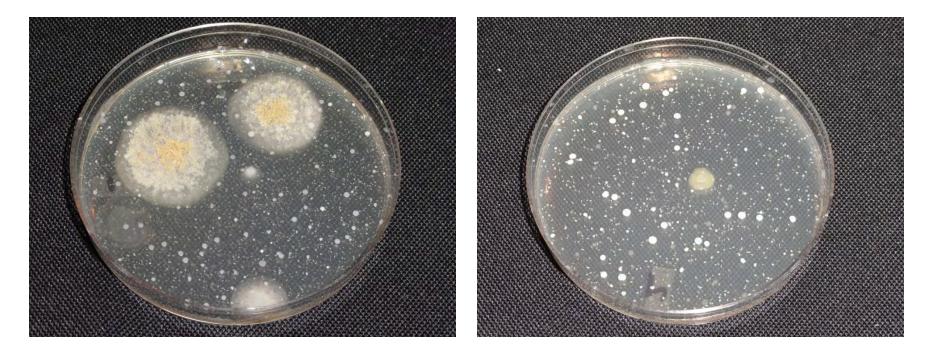








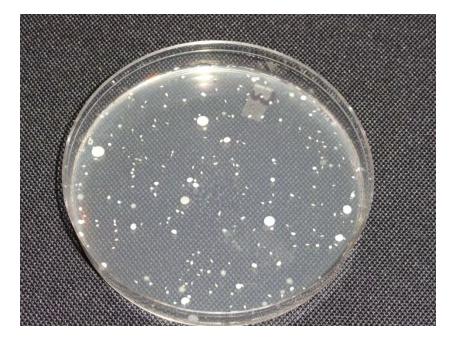
Pour Plate System

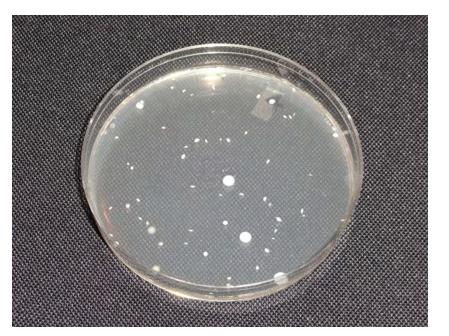


First Sample

Second Sample

Pour Plate System



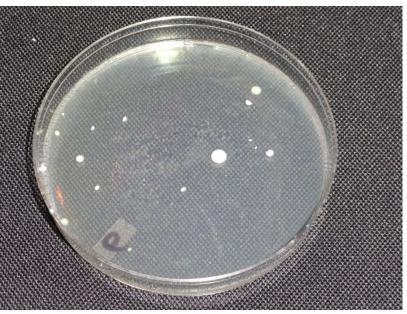


Third Sample

Forth Sample

Pour Plate System





Fifth Sample

Sixth Sample

Pour Plate System



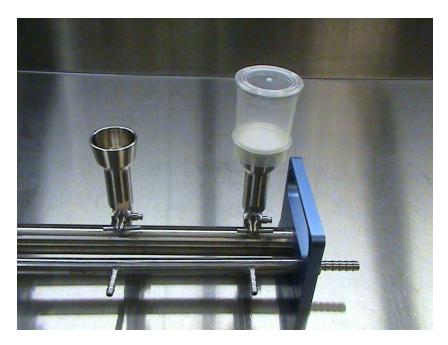
Seventh Sample Note growth around the edge of swab

Liquid Method

- The broth can be processed as follows
 - Membrane Filtration
 - Spread plate

Membrane Filtration



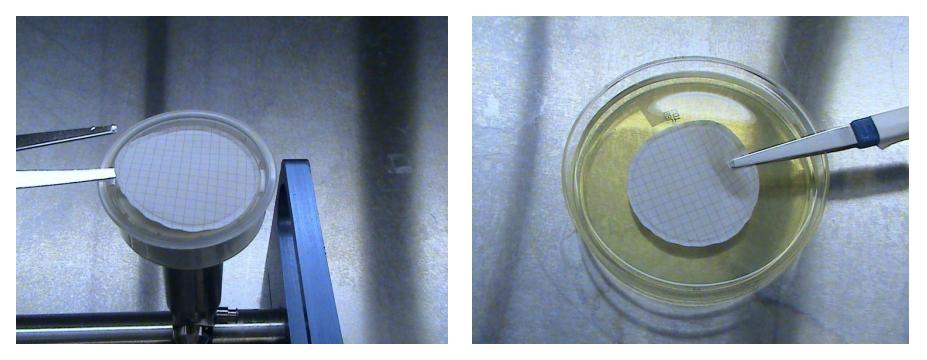


Membrane Filtration

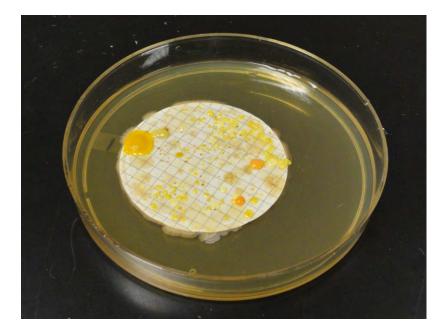


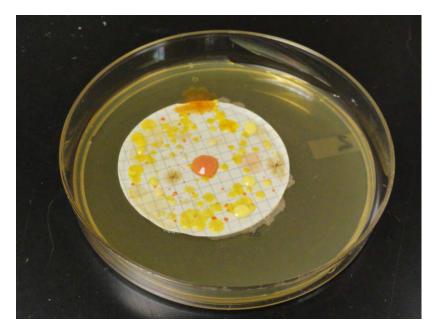


Membrane Filtration



Membrane Filtration

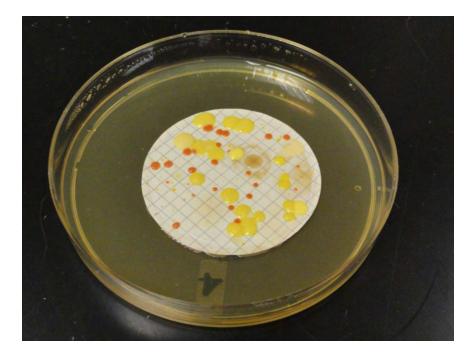


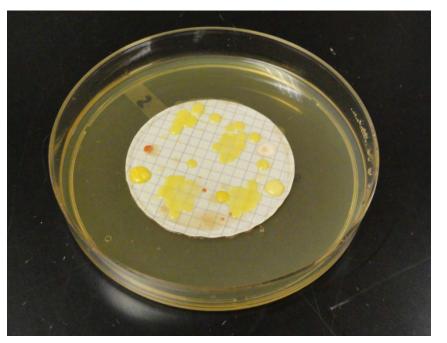


First Sample

Second Sample

Membrane Filtration

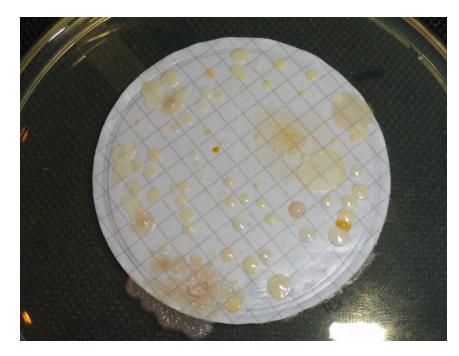


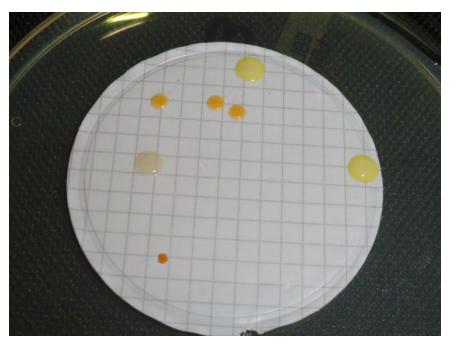


Third Sample

Forth Sample

Membrane Filtration

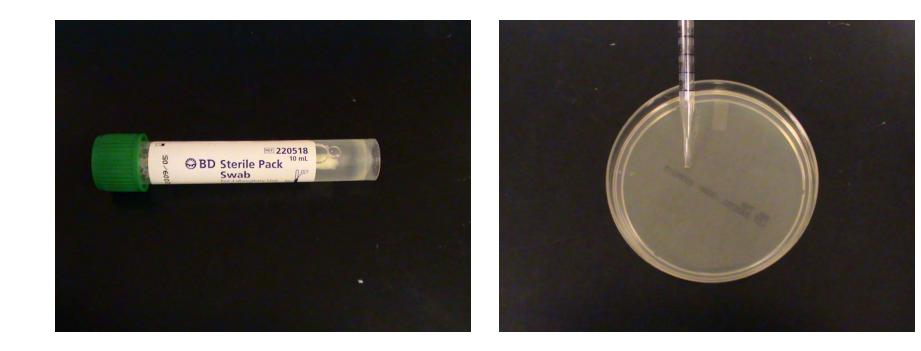




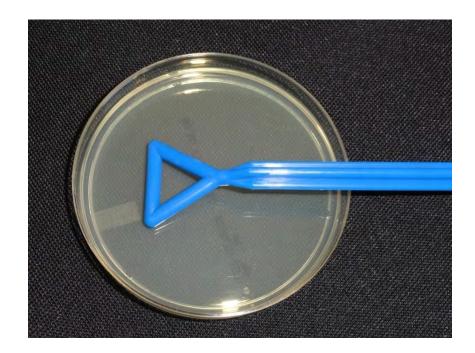
Fifth Sample

Sixth Sample

Spread Plate System



Spread Plate System



Spread Plate System



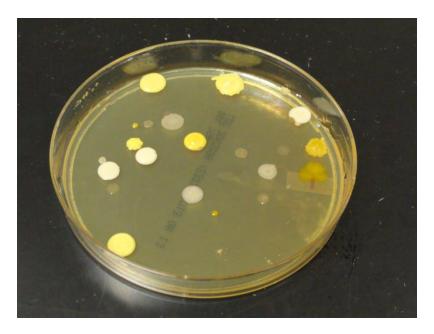


First Sample

Second Sample

Spread Plate System

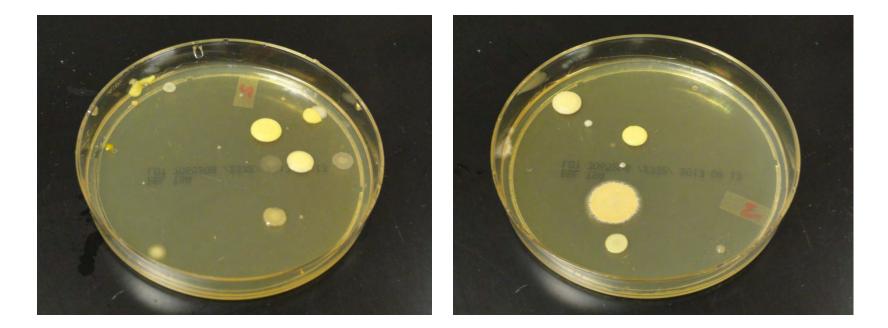




Third Sample

Fourth Sample

Spread Plate System



Fifth Sample

Sixth Sample

Swab Testing for Irregular Surfaces

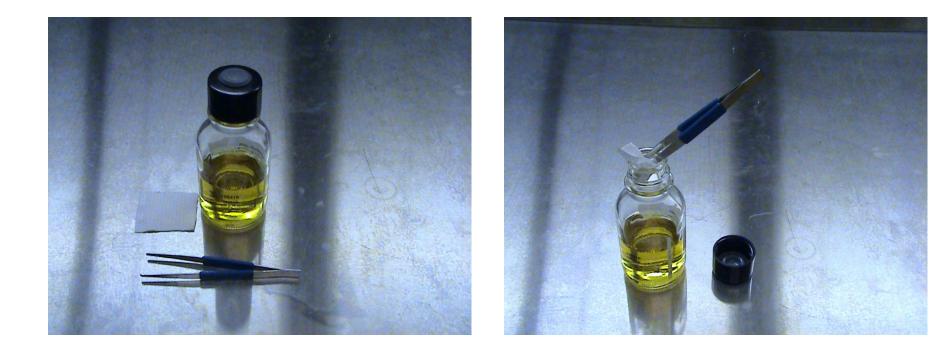
- Advantages
 - Useful for irregular surfaces
 - Can be used on flat surfaces

- Disadvantages
 - Sampling techniques could effect results
 - Requires manipulation to culture sample
 - Time consuming
 - Variable based on operator techniques
 - Recovery rate very low

Surface Wipe Method

- Wipe the testing area with a sterile wet wipe to pick up any CFU's present
- Place sterile wipe into TSB
- Incubate

Surface Wipe Method



Surface Wipe Method



Advantages

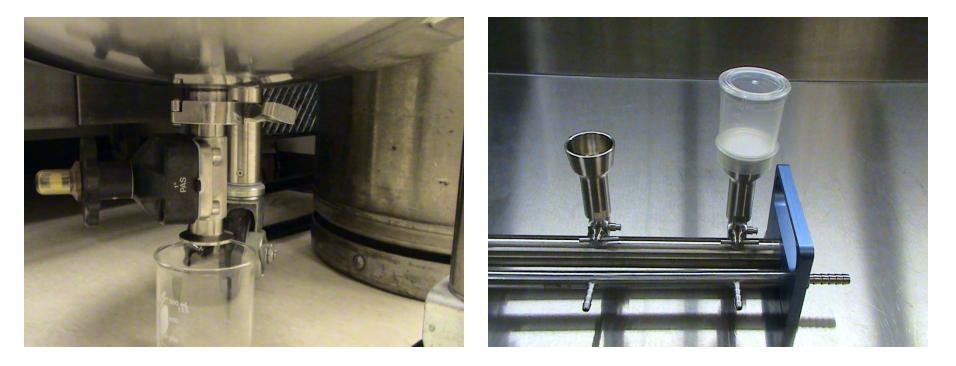
- Easy to use
- Can sample a large surface area
- Sample flat and irregular surfaces

- Disadvantages
 - Can get messy
 - Requires a large volume of media in the sampling area
 - Media can tip over
 - Placing sterile wipe into the media can be problematic

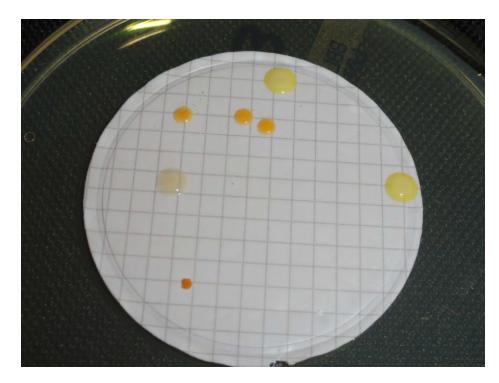
Surface Rinse Method

- In this method, the container/surface is rinsed with water
- The rinse water is membrane filtered
- The filter is placed on a agar surface
- Used mainly for interior of tanks

Surface Rinse Method



Surface Rinse Method



Advantages

- Useful for large surface areas, where interior surface bioburden is needed.
- Some examples are kettles, tanks and equipment.
- Useful for irregular surfaces.

- Disadvantages
 - Not suitable for most applications.
 - Requires extensive manipulations.
 - Techniques and sample processing can affect results.

Yeast and Mold Testing

- Can be combined with bacterial monitoring
- Performed with routine monitoring or at a minimum quarterly
- > Take a second sample using selective agar
 - Sabaraud Dextrose Agar (SDA)
 - Potato Dextrose Agar (PDA)
 - Rose Bengal Agar (RBA)
- Incubation 20 25 °C

Incubation Times and Temperatures

- Samples should be placed into the incubator in less than 4 hours after sampling
 - This time needs to be qualified
- The lower temperatures require longer incubation
- Incubation duration between 48 168 hours

Incubation Times and Temperatures

- Incubation temperatures

- Anaerobic Monitoring
- Media Types and Vendors
- Contamination Control
- Qualification/Validation of Methods

Takeaway Message

- Ensure proper viable air sampler is being used based on exhaust air
- Confirm all sample locations are justified based on the validation data
- Routing sampling is not performed in static conditions
- Ensure all testing methods are optimized