

### **Interactive Session:**

Designing a virus filtration process - assumption and points to consider, How to design a process, Calculating production costs

Dr. Sebastian Teitz, Product Manager & Scientific Coordinator, Asahi Kasei Bioprocess Europe, s.teitz@akbio.eu, www.ak-bio.com



In the following slides we will discuss filtration process:

- related to **Filterability** (Flux, Volume)
- but NOT related to virus removal (LRV)

The presentation will give guidelines and strategical approaches, but cannot guarantee the performances for the manufacturers particular process and protein-solutions



### **Vocabulary:**

**TransMembrane Pressure (TMP):**  $P_{Feed} - P_{Filtrate} = kPa$ 

**Flow rate:** Filtration Volume / min or h = L/h or L/min

Flux: « Flow rate / Filtration surface area» = L/h/m<sup>2</sup> or LMH

Filtration Capacity: « Volume / Filtration surface area» = L/m<sup>2</sup>

Mass Throughput: « g of target protein / Filtration surface area» = g or kg /  $m^2$ 

Before the filter: Feed solution After the filter: Permeate or Filtrate solution



Main equations:

```
S (filtration surface area) = Volume of protein solution / (Flux x Time)

or

S (filtration surface area) = Volume of protein solution / (Filtration Capacity)
```

- Why filtration flux & capacity important?
  - To reduce the filtration surface area (S)
  - To reduce filtration time
  - To reduce cost!



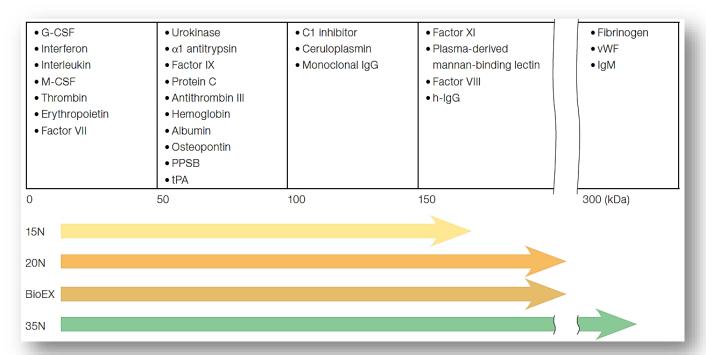
### Preliminary information required (if known):

- Plasma derived product or product from cell culture?
- Protein type (mAb, Fab, clotting factor, etc..), sensitivity to temperature & other stress
- Molecular weight
- Buffer composition, concentrations, pH and conductivity
- Concentration of protein prior to nanofiltration step
- Commercial batch volume to be nanofiltered at once
- Maximum acceptable time for nanofiltration
- Purification steps prior to nanofiltration
- Target virus to remove, expected removal (LRV)
- Other removal/inactivation steps
- Availability of fresh solution for lab scale trials
- Single use or SS equipment



- Preselection of the nanofilter (pore size, parvovirus/retrovirus removal):
  - ✓ What is the size of my target protein ?
  - Can I use a parvovirus removal filter grade?

Example: easier to nanofilter Fibrinogen with Planova 35N vs. 20N





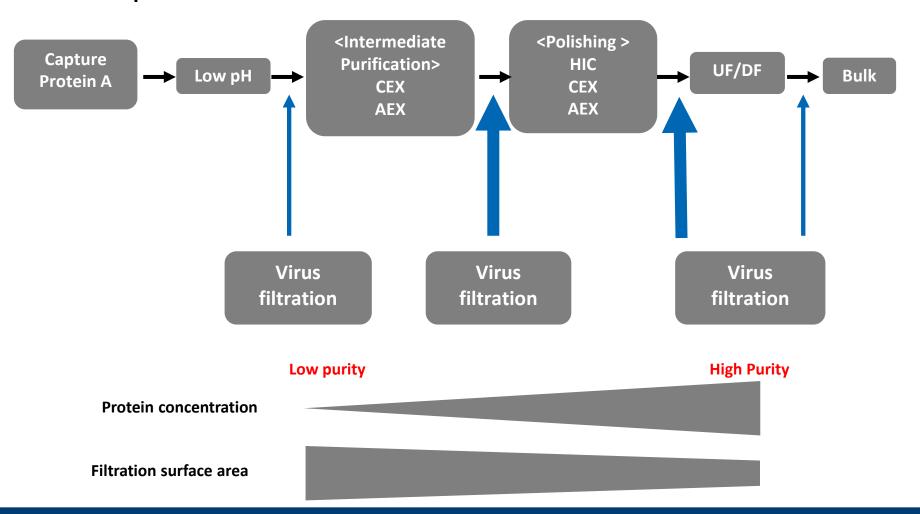
### What protein concentration ?

- Always an optimal concentration to get the maximum mass throughput (g or kg /m²)
- ✓ The higher the concentration, the higher the mass throughput until a certain point, then decline
- ✓ Main reason: Polarization Layer (= "traffic jam")





What protein concentration ?



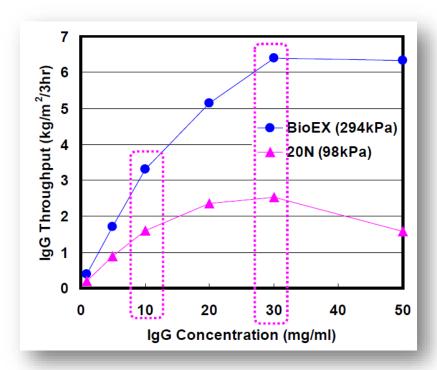


### Which protein concentration ?

Solution: h-IgG (polyclonal) in 100 mM NaCl, pH 4.5

Filtration pressure: 79 kPa (20N), 294 kPa (BioEX) Filtration

time: 3 h



- ✓ Could be different with another product, buffer or operating conditions
- ✓ Optimal concentration ~30 g/L

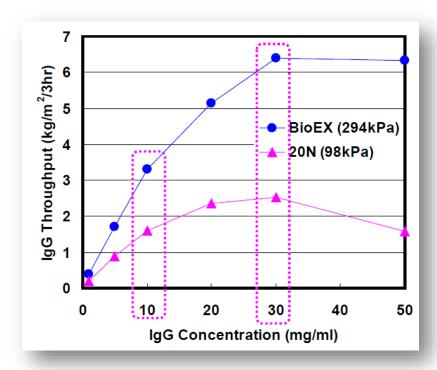


### Which protein concentration ?

Solution: h-IgG (polyclonal) in 100 mM NaCl, pH 4.5

Filtration pressure: 79 kPa (20N), 294 kPa (BioEX) Filtration

time: 3 h



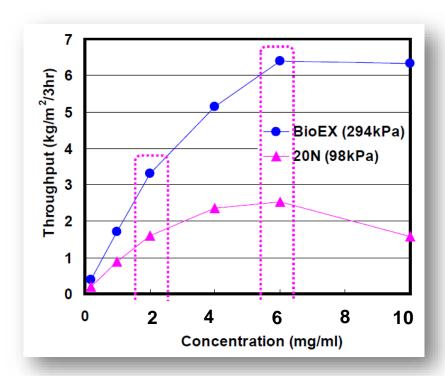
### 10 kg lgG to nanofilter:

- ✓ at 10 mg/mL:
  - 20N: 1.5 kg/m<sup>2</sup>  $\rightarrow$  6.7 m<sup>2</sup>
  - BioEX:  $3.5 \text{ kg/m}^2 \rightarrow 2.9 \text{ m}^2$
- ✓ at 30 mg/mL:
  - 20N: 2.5 kg/m<sup>2</sup>  $\rightarrow$  4 m<sup>2</sup>
  - BioEX:  $6.5 \text{ kg/m}^2 \rightarrow 1.5 \text{ m}^2$



### What protein concentration ?

- Could be different with another product, buffer or operating conditions
- ✓ Sometime it is needed to dilute to lower the protein concentration!

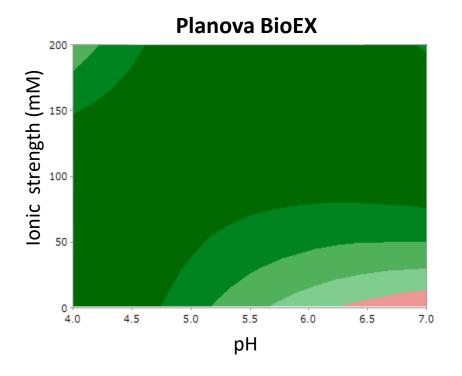




### What pH and Conductivity?

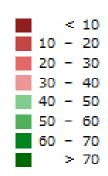
Product solution: 10 mg/mL h-IgG

Filtration time: 3 h



Relative flux (%) =  $(3 \text{ h flux / initial flux}) \times 100$ 

### **Relative flux**



Hongo-Hirasaki, Asahi Kasei Medical, *Planova Workshop*, Prague, 2017 (adapted)



### What prefiltration?

	High Flux Nanofilter	High Capacity Nanofilter
Brand	VPro (Merck) Virosart CPV & HF (Sartorius), Pegasus Prime (Pall)	Virosart HC (Sartorius), Pegasus SV4 & DV20 (Pall), Planova 20N & BioEX (Asahi)
Prefiltration	Special or adsorptive prefilters	None or simple 0.2 or 0.1µm

- ✓ Main parameter impacting flux or filtration capacity → aggregate content
- ✓ Other process-specific contaminants possible
- ✓ In case of very difficult filterability → Prefilter



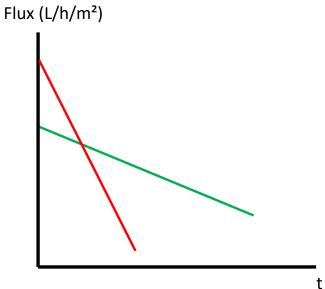
### What Transmembrane Pressure (TMP)?

- ✓ To follow the recommendations of each nanofilter supplier
- ✓ To anticipate Manufacturing constraints (max pressure, equipment)
- ✓ Increase of TMP can improve flux (= lower filtration area) if "reasonable" flux decay. From 80 kPa to 98 kPa → + 22.5% water flux
- ✓ With a too prominent flux decay, or too high impurity-levels, or high Molecular Weight protein → Decrease TMP!



#### What filtration time?

- ✓ To anticipate Manufacturing constraints (working time shift, etc...)
- ✓ The shorter not always the better in terms of cost effectiveness! Calculate the gain of longer filtration time on filtration surface area



For the green curve, extending filtration time can be very beneficial



### What DoE for preliminary trials?

- ✓ Use the experience of the nanofilter supplier!
- ✓ As shown until now, many parameters can impact on the nanofiltration performances
- ✓ Strategy:
  - 1) Understand your product: what protein, MW, pl, pressure or temp sensitive, etc...
  - **2) Understand your process conditions & constraints:** buffer, max desired filtration time, max possible pressure, place of the nanofiltration in the process strain, etc...
  - 3) Preliminary DOE with a few variables and using current buffer
  - 4) Results of the preliminary experiments → "optimized" DOE



### What DoE for preliminary trials?

### **Example:**

- ✓ Product: ~ 100 kD, no temp or pressure sensitive, tendency to aggregation
- ✓ Process: no specific process constraints, 6 h filtration time max
- ✓ Preliminary DOE:
  - 1) Nanofilters: 20N, BioEX
  - **2) Prefilter:** standard 0.1 μm (off-line is ok)
  - **3) TMP:** 50 & 98 kPa for 20N; 150 & 300 kPa for BioEX
  - 4) Conc.: before & after last chrom., before & after last UF/DF
  - **5) Product:** fresh (if possible)
  - **Buffers:** as currently in the different process steps
  - **7) Filtration time:** 6 h, at least 3 h



What DoE for preliminary trials?

### **Example:**

- ✓ Product: ~ 100 kD, no temp or pressure sensitive, tendency to aggregation
- ✓ Process: no specific process constraints, 6 h filtration time max
- ✓ Preliminary DOE: limited volume available or no time...
  - 1) Nanofilters: 20N, BioEX
  - **2) Prefilter:** standard 0.1 μm (off-line is ok)
  - 3) TMP: 50 & 98 kPa for 20N; 150 & 300 kPa for BioEX
  - 4) Conc.: before & after last chrom., before & after last UF/DF
  - **5) Product:** fresh (if possible)
  - **Buffers:** as currently in the different process steps
  - 7) Filtration time: 6 h, at least 3 h



Reality on the field can be different... "evolving" DoE

Solution: monoclonal IgG 1, 24 mg/mL, 10 mM NaCl, pH 5.5

Filtration pressure: 98 kPa (20N-0.001m<sup>2</sup>); 294 kPa (BioEX-0.001m<sup>2</sup>)

0.1 µm prefiltration

**Objectif:** use of BioEX to filter high concentration solution...

#### Test 1:

Filter	BioEx
Product concentration (mg/mL)	24
Buffer	pH 5.5
Total filtration time (min) :	15
Total filtration volume (mL) :	0,6
Average flux (L/h/m2):	2,4
Pressure (bar) :	2,0
Vmax (L/m2):	1,5

✓ It seems it will not be so easy



Reality on the field can be different... "evolving" DoE

#### Test 2:

Filter	BioEx
Product concentration (mg/mL)	12
Buffer	pH 5.5
Total filtration time (min):	38
Total filtration volume (mL) :	7,8
Average flux (L/h/m2):	12,3
Pressure (bar) :	2,0
Vmax (L/m2):	25

#### Test 3:

Filter	P20N
Product concentration (mg/mL)	12
Buffer	pH 5.5
Total filtration time (min):	65
Total filtration volume (mL):	4,0
Average flux (L/h/m2):	3,7
Pressure (bar) :	0,8
Vmax (L/m2):	9

- ✓ First action:

  decrease protein

  concentration.
  - ... but no significant positive impact
- Even 20N cannot nanofilter the solution



Reality on the field can be different... "evolving" DoE



Filter	P20N	
Product concentration (mg/mL)	12	
Buffer	pH 6.6 - 60	mM NaCl
Total filtration time (min):	35	
Total filtration volume (mL):	22,8	
Average flux (L/h/m2):	39,0	
Pressure (bar) :	0,8	
Vmax (L/m2):	> 500	

#### Test 5:

Filter	P20N	
Product concentration (mg/mL)	12	
Buffer	pH 5.5 - 10	0 mM NaCl
Total filtration time (min) :	30	
Total filtration volume (mL) :	20,1	
Average flux (L/h/m2):	40,2	
Pressure (bar) :	0,8	
Vmax (L/m2):	> 800	

- Second action: change pH and conductivity
- Higher
   conductivity has
   a significant
   positive impact
   (High Vmax)



Reality on the field can be different... "evolving" DoE

#### Test 6:

Filter	P20N	
Product concentration (mg/mL)	24	
Buffer	pH 5.5 - 10	0 mM NaCl
Total filtration time (min):	10	
Total filtration volume (mL):	1,3	
Average flux (L/h/m2):	7,5	
Pressure (bar) :	0,8	
Vmax (L/m2):	3,15	

#### Test 7:

Filter	BioEx	
Product concentration (mg/mL)	24	
Buffer	pH 5.5 - 10	0 mM NaCl
Total filtration time (min):	35	
Total filtration volume (mL) :	30,6	
Average flux (L/h/m2):	52,4	
Pressure (bar) :	2 - 3	
Vmax (L/m2):	> 300	

- ✓ Third action: back to the initial high concentration
- ✓ Concentration too high for 20N but BioEX works well !



Data Analysis (Filtration Study Report, Estimation L/m², Vmax)

#### Asahi KASEI

BIOPROCESS

Planova Filtration Report - Study n° XX/XXX/XX

Confidential

#### 1. Objective

The following filtration study took place on the  $26^{th}$  and  $27^{th}$  of January 2016 at X. It was carried out using Planova<sup>TM</sup> 20N and Planova<sup>TM</sup> BioEX filters.

The 3 main objectives were 1) to compare the performances of Planova<sup>™</sup> 20N and Planova<sup>™</sup> BioEX filters at lab scale, 2) to evaluate the impact of a dilution on the filterability of the plasma derived IVIG solution at lab scale, and 3) to perform nanofiltration runs at pilot scale.

#### 3. Manufacturing conditions

Protein: IVIG (160 kD)

Concentration (total proteins): ~ 33 g/L; possibility to dilute down to ~25 g/L

Buffer/Washing solution: 10 mM NaCl, 275 mM Glycine, pH 4.136, Conductivity: 1.6 mS/cm

Volume to nanofilter: 810 L or 1070 L at respectively 33 g/L and 25 g/L (total proteins)

Acceptable nanofiltration time: 6 h maximum



Data Analysis (Filtration Study Report, Estimation L/m², Vmax)

### 4. Experimental materials

Planova<sup>TM</sup> filters: 20N (0.001 m<sup>2</sup> and 0.12 m<sup>2</sup>) and BioEX (0.001 m<sup>2</sup>)

Pre-filter(s): 0.2/0.1 µm (Pall Fluorodyne II Mini Kleenpak)

Protein solution: ~ 3 L were prepared by X (each day) from their pilot scale

production process

Total proteins concentration: Day 1: 32.61 g/L; 25.22 g/L (after dilution)

Day 2: 33.53 g/L; 26.39 g/L (after dilution)

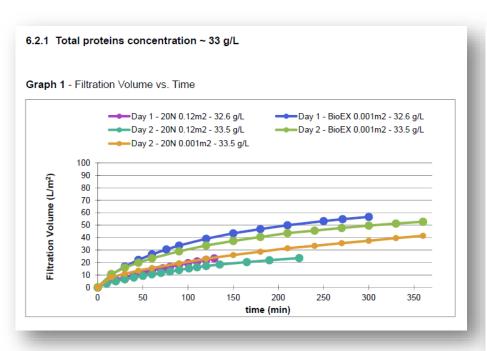


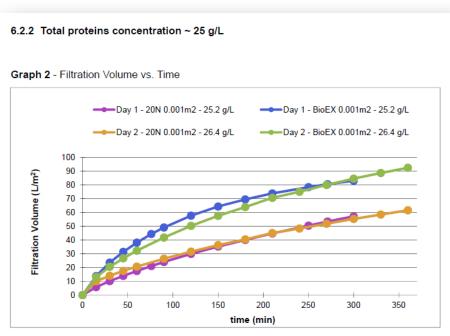






Data Analysis (Filtration Study Report, Estimation L/m², Vmax)





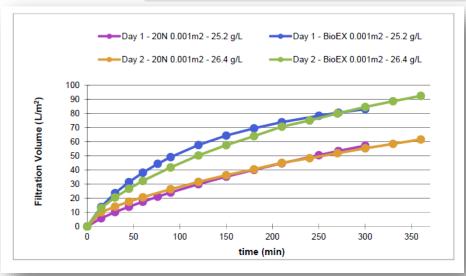
High consistency and scalability of the performances

Positive impact of the dilution on the Filtration Capacity (L/m²)



Data Analysis (Filtration Study Report, Estimation L/m², Vmax)

Total proteins conc. (g/L)	Volume (L)	Filter type	TMP (kPa)	Filter Configuration	Filtration Area (m²)	Filtration Volume (L/m²)	Filtration Time (h)
25	1070			77????	<b>?</b> ??		



#### Filter range:

20N-1m<sup>2</sup>, 20N-4m2 BioEX-1m2, BioEX-4m2

#### Filter cost (example only):

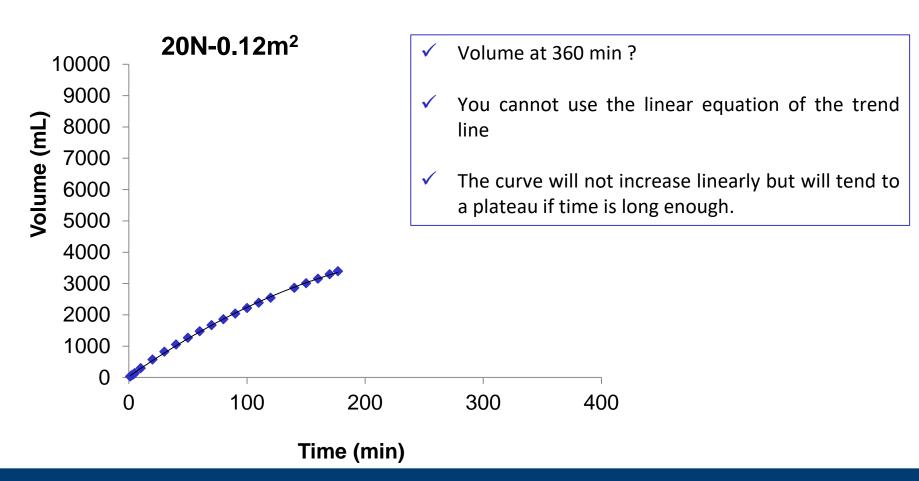
20N: 3000 €/m2 BioEX: 6000 €/m2



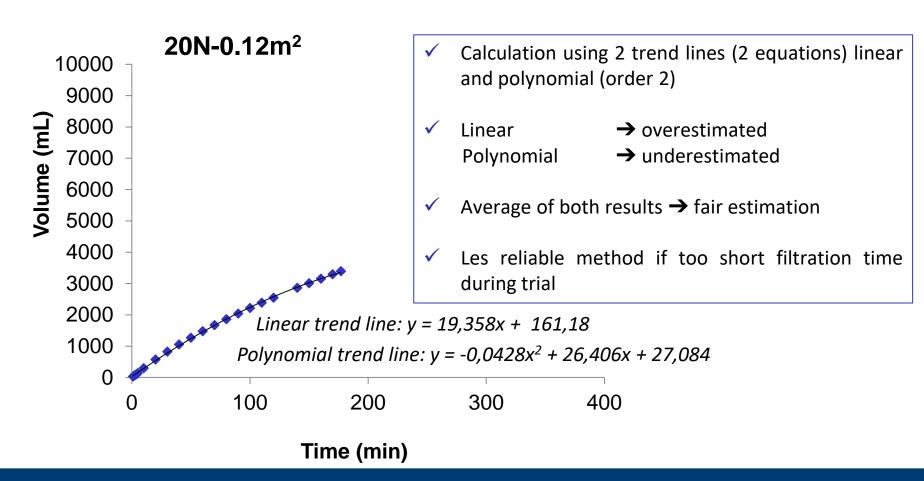
Total proteins conc. (g/L)	Volume (L)	Filter type	TMP (kPa)	Filter Configuration	Filtration Area (m²)	Filtration Volume (L/m²)	Filtration Time (h)
		20N	98	5 filters 20N-4.0 m²	20	54	5
25	1070	BioEX	300	3 filters BioEX-4.0 m²	12	89	6
		BioEX	300	4 filters BioEX-4.0 m²	16	67	3,5

- 1. "Fix" the filter configuration based on the product ranges available → Total filtration surface area (m²)
- 2. Nanofiltration Volume / Total filtration surface area  $\rightarrow$  Filtration capacity (L/m<sup>2</sup>)
- 3. 2 situations:
  - o if the Filtration Capacity was already reached during the trial run you can directly pinpoint the corresponding filtration time.
  - o if you stopped the trial run before achieving the needed Filtration Capacity, you will have to estimate the filtration time (next slides).

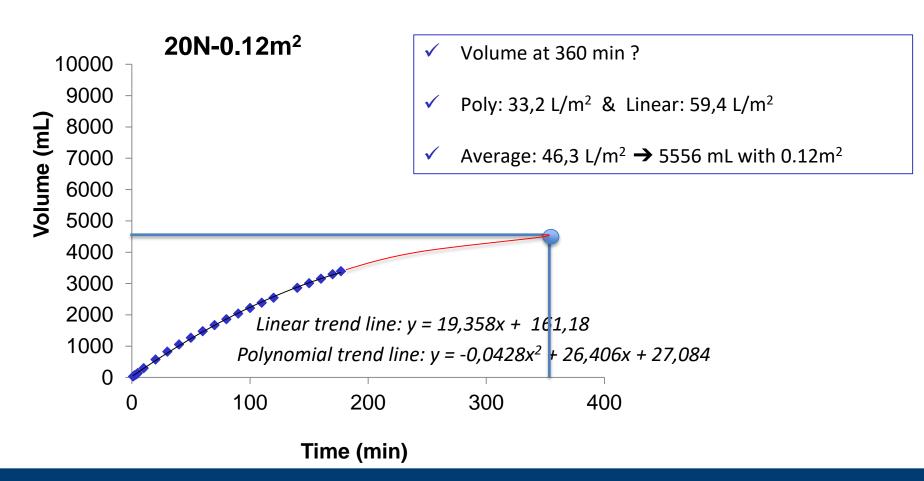














Data Analysis (Filtration Study Report, Estimation L/m², Vmax)

$$\frac{F_{min}}{V} = \frac{1}{V_{max}} + \frac{1}{R \times t}$$

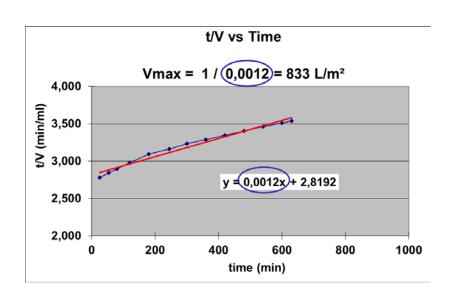
$$F = filtration area (mm^2)$$
  $V_{max} = maximum capacity (l/m^2)$   $V = process volume (l)$   $t = process time (min)$   $R = initial volumetric flow rate (l/m^2min)$ 

#### Translation...

"Vmax is the filtration capacity  $(L/m^2)$  achieved if the time would be infinite"



t (min)	Volume (mL)	dV/dt (mL/min)	/ t/V	Flux (LMH)
0,0	0,00	-	-	-
25	9,00	0,36	2,778	21,60
54	19,00	0,34	2,842	20,69
80	27,63	0,33	2,895	19,92
120	40,30	0,32	2,978	19,01
180	58,16	0,30	3,095	17,86
245	77,52	0,30	3,160	17,87
300	92,75	0,28	3,235	16,61
360	109,50	0,28	3,288	16,75
420	125,50	0,27	3,347	16,00
480	141,00	0,26	3,404	15,50
540	156,00	0,25	3,462	15,00
600	171,00	0,25	3,509	15,00
630	178,10	0,24	3,537	14,20



- ✓ The main interest of Vmax is to inform about the possibility to filter longer and then to achieve high filtration capacity
- ✓ The higher Vmax, the lower the flux decay



Exercise! ©

→ Calculation of Production Costs



# **Questions?**

