Thermal treatment of Hyaluronic Acid Pre-filled Syringes: challenges to overcome *Maria Luisa Bernuzzi, Fedegari Autoclavi SpA*

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Open discussion

- •What have we to sterilize?
- •What have we to look at?
- •What are the «critical parts»?
- •What are the risks?

What we have to sterilize **The product**

- First study of PFS with Sodium Hyaluronate (M.L. Bernuzzi, A. Giori): a review of it
- Sterilizing treatment based on temperature control
- Sterilizing treatment based on F_0 control
- Many applications for hyaluronic acid
- Counterpressure autoclaves
- What happens when an aqueous solution in a sealed container is heated

Hyaluronan

- Linear polysaccharide
- Repeating N-acetyl-D-glucosamine and D-glucuronic acid units
- High molecular mass
- Viscoelastic properties

Hyaluronic acid degradation

- Several sterilization methods:
	- *Microwaves*
	- *UV*
	- *Gamma rays*
	- *High Temperature*

The viscosity of the solutions decreases in time as a function of temperature

An optimized moist heat sterilization treatment can minimize the HA degradation

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The study

Proving that a well-developed moist heat sterilization cycle can minimize HA degradation

Sterilization treatment

Three sterilization treatments were performed on Sodium Hyaluronate prefilled syringes:

The sterilization process was controlled by F_0 *target = 15' using the Kaye validator.*

Sodium hyaluronate PFS

- HA molecular weight = 1.6×10^6 Dalton
- Solution formulation: polysaccharide hydrated in a physiological phosphate buffer solution for 12 hours at 50 °C
- The solution was divided in glass prefilled syringes (PFS), containing 50 mg/2,5 ml of Sodium Hyaluronate

Moist heat sterilization

It is the method of choice used for the sterilization of PFS

«Sterilization by saturated steam under pressure is preferred, wherever applicable, especially for aqueous preparations».

European Pharmacopeia 10th edition

Physics of a syringe during a sterilization cycle

Increase of the internal pressure inside the syringes during the heating phase due to:

- 1. Water evaporation
- 2. Dissolved gases that leave the solution
- 3. Thermal expansion of the liquid
- 4. Air present in the head space

Counter-pressure steam sterilization

To balance the overpressure inside the syringes and to reduce the risk of plunger expulsion:

The air pressure contained in the chamber is not extracted but it is "controlled" during the entire cycle Obviously, the chamber air pressure increases with the heating The total pressure in the chamber is given by the partial

pressure of the heated air plus the partial steam pressure

Counter-pressure

Pressure inside the chamber at the different temperatures used in the cycles performed on PFS

Syringe without head space

Both in the case of **glass and polycarbonate**, the pressure required to prevent any temporary movement of the plunger in a completely filled PFS exceeds by a factor of **one hundred** the pressure which can be applied in a moist-heat sterilizer. The above is coherent with both the common sense (liquids are typically "incompressible" but under very high pressure)! **Polyethylene and polypropylene** containers could fully compensate the thermal expansion of water, but both mechanical resistance and chemical stability of them depend a lot on the specific qualities of these plastics and they may be unsuitable for syringes to be exposed at 121 °C.

Enough length must be always provided at the open end of a glass or polycarbonate PFS to allow the plunger to "trip outwards" during the sterilization without ever protruding from the syringe. For treatment at 121 to 124 °C, the free length beyond the plunger must be at least 5.8% of the water length inside a glass PFS, and at least 4.2% in a polycarbonate.

F_0 calculation

$$
F_0 = \sum \Delta t \; 10^{\frac{T-121}{Z}}
$$

- Algorithm used to measure the lethality of a thermal sterilization process
	- It calculates, by measuring physical parameters (time and T)
	- the amount of heat (lethal dose) delivered to the product

- Δt = time interval between subsequent *temperature measurements*
- *T= actual sterilization temperature*
- *z*= temperature coefficient (10 °C for F₀ calculation)

F_0 calculation

- F⁰ target = **15 minutes**
- Beginning of F_0 accumulation at 111 °C
- \cdot F₀ accumulation during heating, sterilization and cooling
- Sterilization phase stopped at $F_0 < 15'$
- Thermocouple device (TC) inserted into a "reference syringe"

Structure of the study

Three sterilization treatments were performed on Sodium Hyaluronate PFS:

Structure of the study

For each temperature tested:

a) Preliminary trial including a PFS with a TC

 F_0 accumulated during heating, sterilization, cooling

b) At least two replicates with 15 PFS

Structure of the study

Pt 100 probes

Temperature probes locations

- \triangleright 1 PFS was vertically attached to the side of the tray inside the machine using zip-ties. The thermocouple was placed inside the solution, ensuring that it was not touching any side of the glass syringe.
- ➢ Other temperature probes (Pt100) were located within the autoclave chamber to additionally monitor the temperature during the sterilization process.
- \triangleright The autoclave temperature probes were free in the chamber.

Loads configuration

• Load type 1

'Home-made' method to insert the termocouple in the syringe

Sample with the TC inserted directly inside the syringe

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Test performed

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Temperature-Time profiles

Temperature-Time profiles

Temperature-Time profiles

130°C peak cycle

- \checkmark Fast heating/cooling of the product
- \checkmark Sterilization at high temperature and short holding time
- ✓ Higher counterpressure

How to know about product degradation

GPC (Gel permeation chromatography)

Useful to know the molecular weight of the product and discover the degradation

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How does GPC work

Rheological investigation

After the cycle, *a rheological investigation* was performed in order to understand if the different processes had damaged the product. Viscosity is a parameter used in order to understand the degradation of the product.

A *«cone on plate rheometer»* was used.

Viscosity analysis

Identification of two main shear rate intervals

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Viscosity analysis

Entanglement and disentanglement.

During shearing a certain number of macromolecules are oriented into shear direction (*dis-entanglements*). As consequence the viscosity tends to decrease in this parts of volume. Simultaneously however other molecules, previously disentangled, are *recoiling and re-entangling* again because of their viscoelastic behavior. At low shear conditions, the overlapping of these effects (dis- entenglements and reentanglements) occurs as a costant limiting value of the viscosity function.

On the other side *at increased shear rate*, the number of *disentanglements* is exceeding the number of re-entanglements: the curve of the viscosity is decreasing continuously.

Dynamic viscosity

Conclusions of the first study

- ❖ Viscosity reduction at the end of all cycles performed.
- ❖ The worst cycle is the one run at 115 °C.
- ❖ The viscosity change is reduced during a **peak cycle**

The higher is the temperature and the lower is the exposure time, the lower is product degradation

Sterilizing treatment based on temperature control

Contrary to what one could expect, the treating at lowest temperature and for longest time leads to significant reduction of polymer degradation and its viscosity.

A peak cycle at high temperature and short time preserves the integrity of the sample.

Sterilizing treatment based on temperature control

Sterilizing treatment based on temperature control

The customer may not have the proper autoclave to reach **130°C** because an over-rating machine is required.

In this case, the sterilization cycle at **125°C** could be selected as the best compromise.

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PFS with sodium hyaluronate: viscosity reduction

Load treatment strategy: viscosity reduction

Possible strategies to treat heat-sensitive loads

- a. To minimize the microbial load of the product to be sterilized, so as to be able to minimize the heat dose (temperature-time) required in order to achieve an adequate SAL (Sterility Assurance Level).
- b. To make the heating/cooling phases as rapid as possible
- c. To raise the sterilization temperature, adequately reducing its duration: this solution can be clearly implemented in combination with solution b.

How to have a further reduction of the product degradation

Since HA is easy to degrade, it's better to approach a sterilizing treatment based on the F₀ control.

An approach with $F0$ equal to $8'$ is tested, in order to try if the degradation could be reduced.

Sterilizing treatment based on F₀ control

KAYE validator is used to control the reached F0. It is connected with a set of termocouples, and one of them is inserted inside a syringe.

This is the best way to know the delay of the heat accumulation caused by the hyaluronic acid.

Viscosity vs Shear rate

Many application for hyaluronic acid

- Ophthalmic
- Cosmetic
- Osteoarthritis therapy

There are many different viscosity grade of hyaluronic acid and it's better to choose according to the proper use.

Differences depending on HA applications

Autoclave (Air Over steam) sterilization of HA

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Choose the best FOA sterilization cycle

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Sterilization cycle with FOW

Sterilization cycles, using an FOW autoclave, were performed, in order to observe any **differences** in the **product degradation**.

Sterilization temperature

Remarks:

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רוציי)

No differences in terms of degradation between **FOW** and **FOA** at different

Cycle performed with a limited load, and

temperatures with same lethality effect

small volume syringes

Counterpressure autoclaves

Steam-air mixture autoclave (FOA)

PFS treatment: which to choose?

Superheated water autoclave (FOW)

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What we have to sterilize?

The container: a syringe with a LUER LOCK

Does a sterilization cycle affect the mechanical characteristic of a syringe?

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Impact of the steam sterilization on the PFS *Does the steam sterilization cycle damage the*

primary packaging (syringe)?

Total Length test

Torque Force test

Pull Off Force test

Unscrewing test

The results of the sterilization process with autoclave at **different temperatures** (115°C, 121°C, 125°C, 130°C) are compared to the sterilization with Ethylene Oxide (EtO). The **EtO treatment** was used in the testing as **control**.

Syringes maintain the characteristics required by the manufacturer's customers

Mechanical characteristic of a syringe

[https://www.youtube.com/watch?v=j1RwnFj](https://www.youtube.com/watch?v=j1RwnFjHAHQ) HAHQ

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The microbiology of a syringe

Key points:

- ❑ we have to sterilize a heat sensitive load
- ❑ we have to reduce its degradation
- ❑ a tailored cycle could be the best choice

HOW TO PROCEED

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Purpose of the study

Determine the **most resistant bioburden isolate** and perform steam D₁₂₁-value of spores suspended in Hyaluronic Acid Injection

Procedures

BIOBURDEN EVALUATION

Genetic identification with RNA sequencing technique of each isolate

Boil test. 1.0 x 10⁴ spores inoculated into tubes and put in heat bath at 95-100°C for 10 min (Chapter <1229.2> USP 43). After heat treatment, two 1 mL aliquots of each isolate will be plated in Petri plates and incubated for >48h. Initial screening

Crop production of the most resistant isolate; approximately 10⁸ of the most resistant organism will be required to perform a Dvalue analysis.

Procedures

BIOBURDEN EVALUATION

One set of **40 syringes containing the most resistant isolate** suspended in Hyaluronic Acid Injection will be prepared.

Population assays will be performed on the syringes. The purpose of these assays is to **verify that the spore concentration in the syringes is stable** for the duration of the study.

D-value determination in the steam B.I.E.R. at 121°C.

Identification of the most resistant isolate

Another way to go

Do the customers really want to do this procedure every time?

It's necessary to find the **Biological Indicator** which adapts better to the customers' product

Procedures

Syringes inoculated with

The inoculated syringes are compared to BIs present in the market with *B.subtilis* or *G. stearothermophilus.*

If these BIs have a D-value > than the syringe, the customer will use them.

An example

- The most resistant microorganism, in the syringe, with HA, has a $D_{121^{\circ}C} = 0.1'$
- Bacillus Subtilis, 10⁶ spores, in the syringe, with HA, has a $D_{121^{\circ}C} = 0.3'$
- Geobacillus Stearothermophilus, 10⁶ spores, in the syringe, with HA, has a $D_{121^{\circ}C}$ = 3,2'
- Geobacillus Stearothermophilus, 10⁴ spores, in the syringe, with HA, has a $D_{121^{\circ}C}$ = 2,8'
- BI in the market, with B. Subtilis, in an ampoule, 10⁶ spores:

$$
D_{121^{\circ}C} = 0.4^{\prime}
$$

BI in the syringe in it

Table 1. Insertion of SterilAmp SASU Biological Indicator in Hyaluronic Acid syringes

Plunger removal with the help of sterile tweezers

Removal of about 0.3 mL of the product and insertion of the Biological Indicator

Insertion of the plunger: the syringe with Biological Indicator inside is ready

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Syringe thermal behavior

Study of the syringe thermal behavior

A preliminary phase of the test session involves the study of the syringe thermal behavior, to understand the lag time, i.e. the time necessary to heat up the product until it reaches the same temperature of the chamber.

In order to perform this test, a sterilization cycle is performed. After the set sterilization temperature (121°C) is reached in the autoclave chamber (read by a temperature control probe), a calculation of the amount of time needed to reach the same temperature in the syringe (read by the TC connected to the KAYE validator) is carried out.

Several trials have been performed; the lag time was equal to 35 ".

F_0 calculated: the minimum required, according to the US supplier of BIs

• We consider:

- ❖ the D value of the BI we increase it a little bit to provide a certain safety level
- \div the lag time of the product to add the contribute of the packaging
- $\bullet \quad N_0 = \text{initial spores population}$
- $\mathbf{\hat{y}}$ N = target of the cycle

Final $F_0 = D_{121}$ -value (log₁₀^{N_0} – log₁₀ N) + lag time

0,3'

 $0.4'$

F_0 calculated: the minimum required, according to the US supplier of BIs

C- \vert **C C+**

Final $F_0 = D_{121}$ -value (log₁₀^{N_0} – log₁₀ N) + lag time

This cycle was performed, two samples with a BI were inserted into the autoclave, a sample with a negative control was also located into the chamber

At the end of the cycle they were removed and incubated

No growth

Thank you

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