



Identification and Characterization of Microorganisms Using Molecular Methods

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Accugenix

- A cGMP compliant service laboratory specializing in bacterial and fungal identification for pharmaceutical manufacturers
- Dedicated to providing the highest quality, fastest turnaround time (same day) and the lowest price
- Allows manufacturers access to the best technology
 - without the time and expense of researching, purchasing, and validating new systems



Overview of Microbial Identification



Methods for Identification of Microorganisms

- **Classical Methods**

- In-house biochemicals
- Manual phenotypic methods (API Strips, BBL etc.)
- Automated phenotypic methods (Vitek)

- **Cellular Fatty Acids** (MIDI Sherlock System)

- **Carbon Source Utilization** (Biolog)

- **Genetic** (MicroSeq, Riboprinter)



Limitations of Identification Methods

- Phenotypic properties (biochemical / carbon source utilization) can be variable, subjective, dependent on growth parameters and health of organism
- Cellular fatty acid profiles change with temperature, age of culture and growth medium
- Some systems require subjective off-line testing such as gram stain, oxidase, coagulase, etc., before determining the appropriate test card
- DNA Sequencing system is not automated, numerous, somewhat complicated lab procedures, and manual interpretation of results requires microbial phylogenetic experience



A Revolution in Bacterial Taxonomy

G. Fox, et. al. 1980. Science. 209:457-463

- Toward a natural system of classification based on phylogenetics rather than taxonomic characters
- In many cases taxonomic characters are not phylogenetically valid- ie morphological characters such as cell shape, mode of cell division, lack of cell wall.....can be misleading



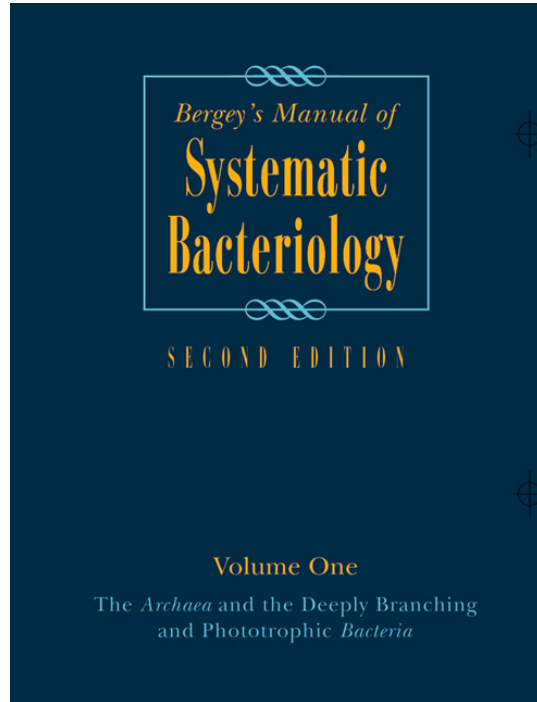
Road Map to Bergey's

George M. Garrity and John G. Holt

"Bergey's Manual of systematic bacteriology, one of the most comprehensive and authoritative works in the field of bacterial taxonomy, has been extensively revised in the form of a five volume second edition. Since the first edition was published in 1984, the field has undergone explosive growth, with over 2,200 new species and 390 new genera having been described. Numerous taxonomic rearrangements and changes in nomenclature have resulted from more than 850 published new combinations.

These developments, which are attributable to rapid advances in molecular sequencing of highly conserved regions of the procaryotic genome, most notably genes coding for the RNA of the small ribosomal subunit, have led to a natural classification that reflects the evolutionary history of Bacteria and Archaea, and to the development of new, universally applicable methods of identifying these organisms."

Phylogenetic Taxonomy



- Universal system
- Reproducible from lab to lab – and over time
- Sequence characters are not subjective
- More informative than phenotypic ID
- No specific growth requirements
- Accepted gold standard for taxonomy



Regulatory Focus



Pharmaceutical cGMPs for the 21st Century

Guidance for Industry **Sterile Drug Products Produced by** **Aseptic Processing — Current Good** **Manufacturing Practice**

DRAFT GUIDANCE

U.S. Department of Health and Human Services
Food and Drug Administration
Center for Drug Evaluation and Research (CDER)
Center for Biologics Evaluation and Research (CBER)
Office of Regulatory Affairs (ORA)

September 2004
Pharmaceutical CGMPs



FDA - Sterile Drug Products Produced by Aseptic Processing

Environmental Monitoring:

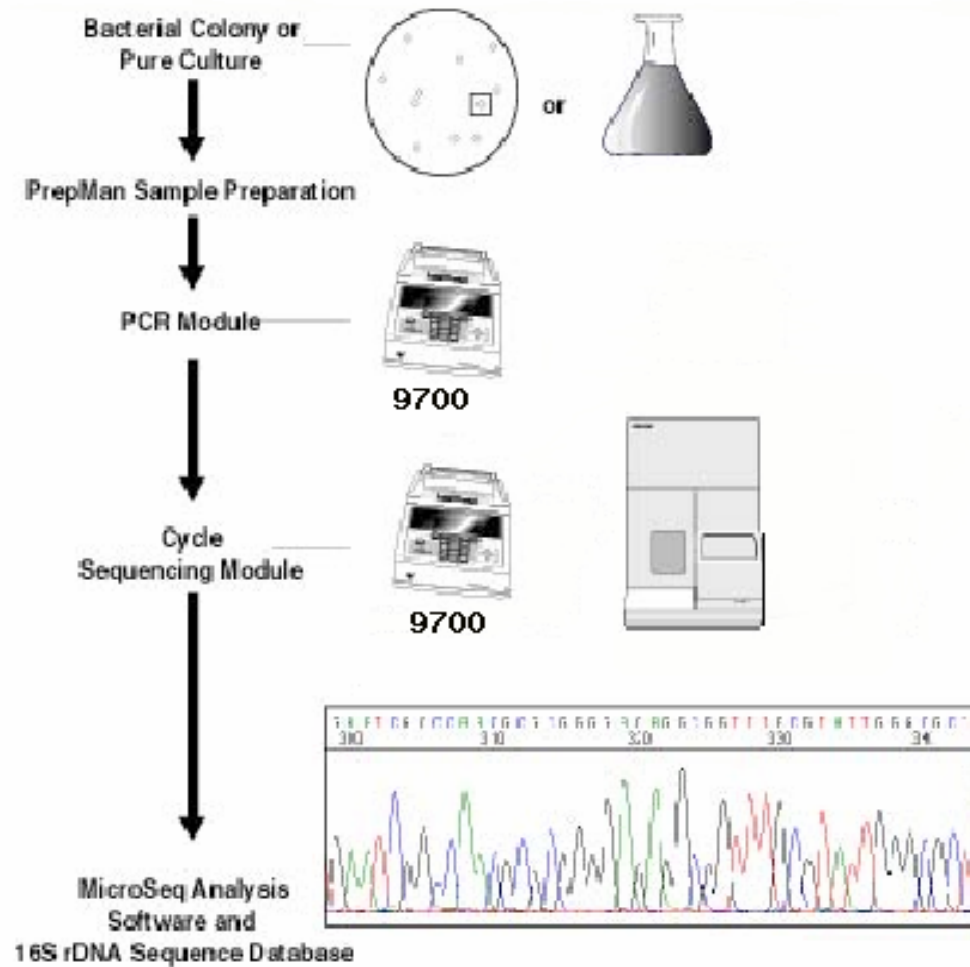
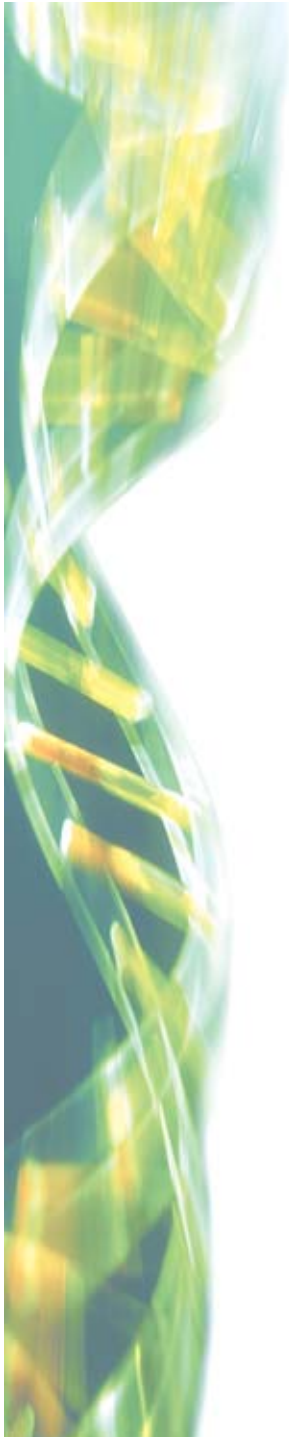
“Genotypic methods have been shown to be more accurate and precise than traditional biochemical and phenotypic techniques. These methods are especially valuable for investigations into failures (e.g., sterility test; media fill contamination).”

Sterility Testing:

“Advanced identification methods (e.g., nucleic-acid based) are valuable for investigational purposes. When comparing results from environmental monitoring and sterility positives, both identifications should be performed using the same methodology”



Accugenix Process





Process

- Sample Accessioning
- DNA Extraction
- PCR
- PCR Cleanup
- Cycle Sequencing
- Cycle Sequencing Cleanup
- Automated Fluorescent DNA Sequencing
- Data Analysis
- Data Interpretation



Sample Accessioning

- Samples are received and checked for damage
- Samples are matched to paper work and check for correct number, match to sample codes, and correctness of requested test code
- Samples are prioritized based on requested Turn Around Time (due date scheduling)
- Samples are accessioned into Master Log and database
- Samples are grouped into batches, and batch records are created

Sample Accessioning

ACCUGENIX
XXXX IDENTIFY WITH THE LEADER

Batch: 50917017
Due Date: Sat 9/17/2005

Type	Lot#	Exp. Date	Thermal-cycler	Sample Sheet	Date/Time	Initial	Run Date	Run Time
DNA Extraction (SOP-GEN-002)	N/A 9/15/05 W	12/10/05		Lot 19460006A Extraction Beads	9/15/05 W		50917017	Sat
DNA Dil. (SOP-GEN-002 / SOP-GEN-012)	1:100 EM 2795B06		9/14/05		9/15/05 W		50917017	Sat
PCR Master Mix (SOP-GEN-004)	F-5001CA (GRO-23)05 WELDS 8				9/15/05 W		50917017	Sat
Batch Order Verification (SOP-GEN-006)							50917017	Sat
PCR Cleanup (SOP-GEN-006)							50917017	Sat
Water (SOP-GEN-006)	EM 2795B06						50917017	Sat
Seq. Master Mix (SOP-GEN-007)							50917017	Sat
Seq. Cleanup Plate (SOP-GEN-010)								
Sequencing Wash / S (SOP-GEN-010)	Seq Wash V94218	05/24/06						
3100 Run Folder (SOP-GEN-027)								

C#	Test Code	Cust Code	Cust ID	Sample Code Verification	Batch Position	Reflux
125415	BacSeq-2	CRN3	#111VPL08/059	S/A 9/15/05	1	
125416	BacSeq-2	CRN3	#112VPL08/065	S/A 9/15/05	2	
125417	BacSeq-2	CRN3	#113VPL08/191	S/A 9/15/05	3	
125418	BacSeq-2	CRN3	#114VPL08/188	S/A 9/15/05	4	
125419	BacSeq-2	CRN3	#115VPL08/187	S/A 9/15/05	5	
125420	BacSeq-2	CRN3	#116VPL08/188	S/A 9/15/05	6	
0	BacSeq-	ACC1	OCNeg	S/A 9/15/05	7	
125421	BacSeq-2	CRN3	#117MED/09/030	S/A 9/15/05	8	

Electropherograms Attached _____
 QC Review _____
 Batch Release _____

SOP-GEN-013.1
 rev. 4/12/2005 jdb

Sample Accessioning

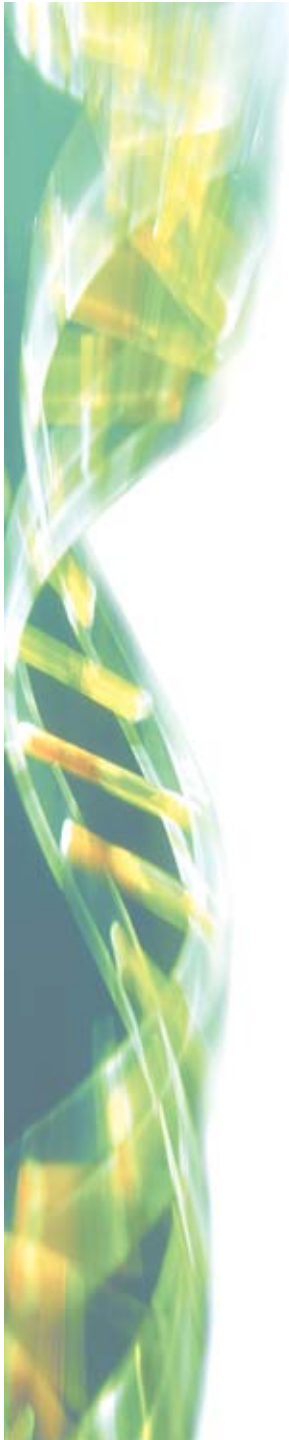




DNA Extraction

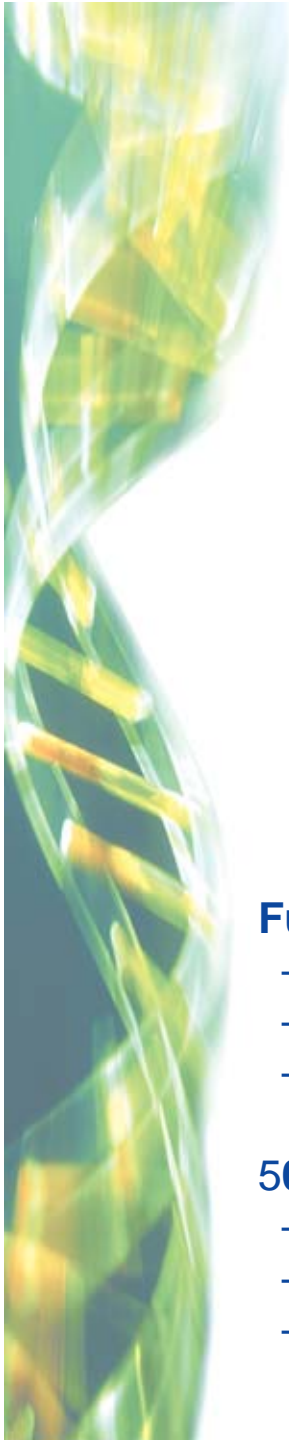
- PrepMan Ultra
 - Cells
 - Paramagnetic Beads
-
- Fast & easy
 - Results in clean DNA
 - Lower failure rate, due to elimination of PCR inhibitors PMU alone does not remove

DNA Extraction



PCR

- Bacterial & Fungal Master Mixes
 - DNA Polymerase
 - dNTP's
 - Primers
- Extracted genomic DNA
- Reactions set-up in a Laminar Flow Hood in a dedicated laboratory to minimize contamination of amplified products
- Dedicated equipment and materials
- Technicians wear disposable gowns while working on PCR set-up



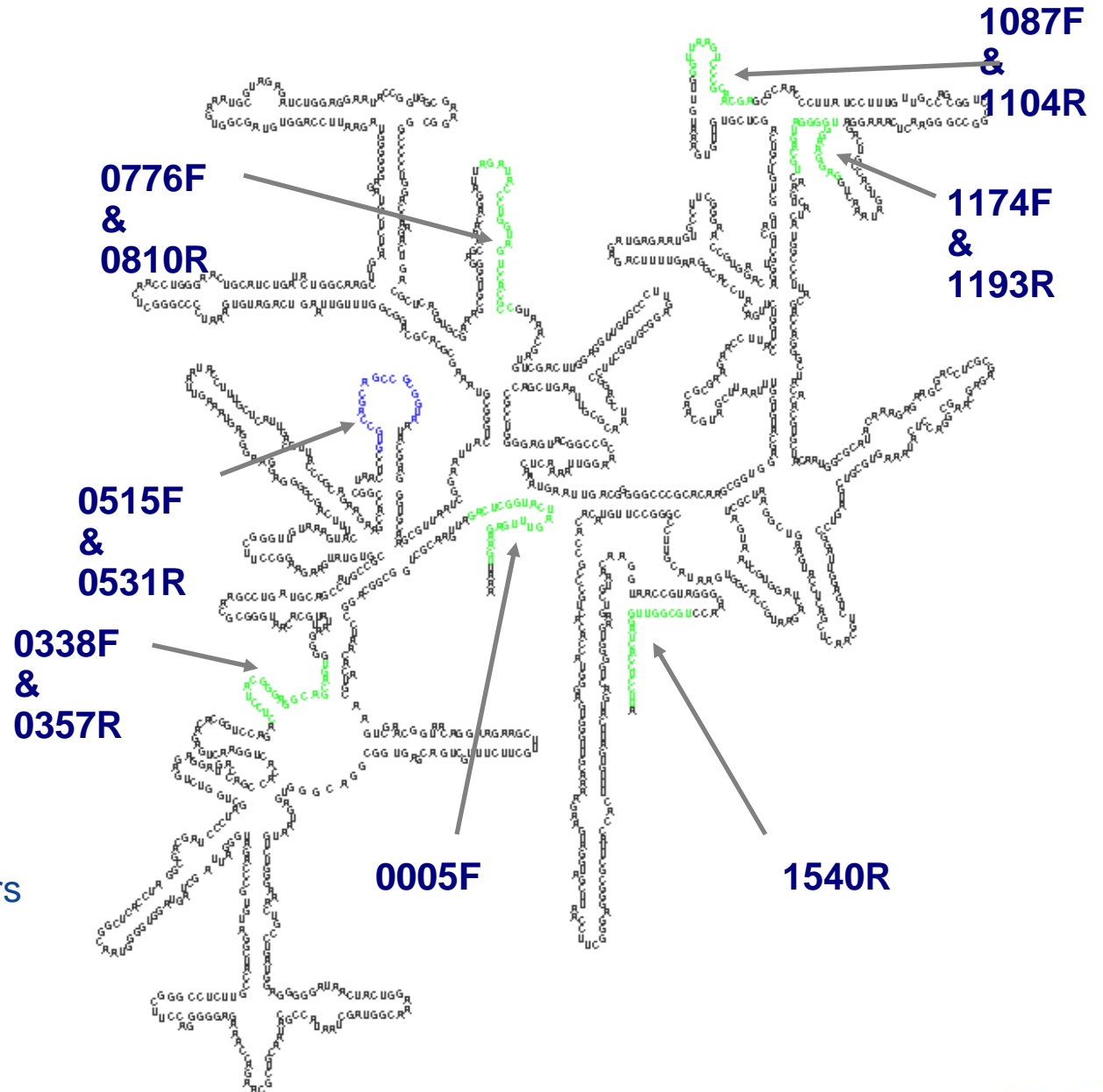
16S rRNA Structure & Primer Locations

Full Gene

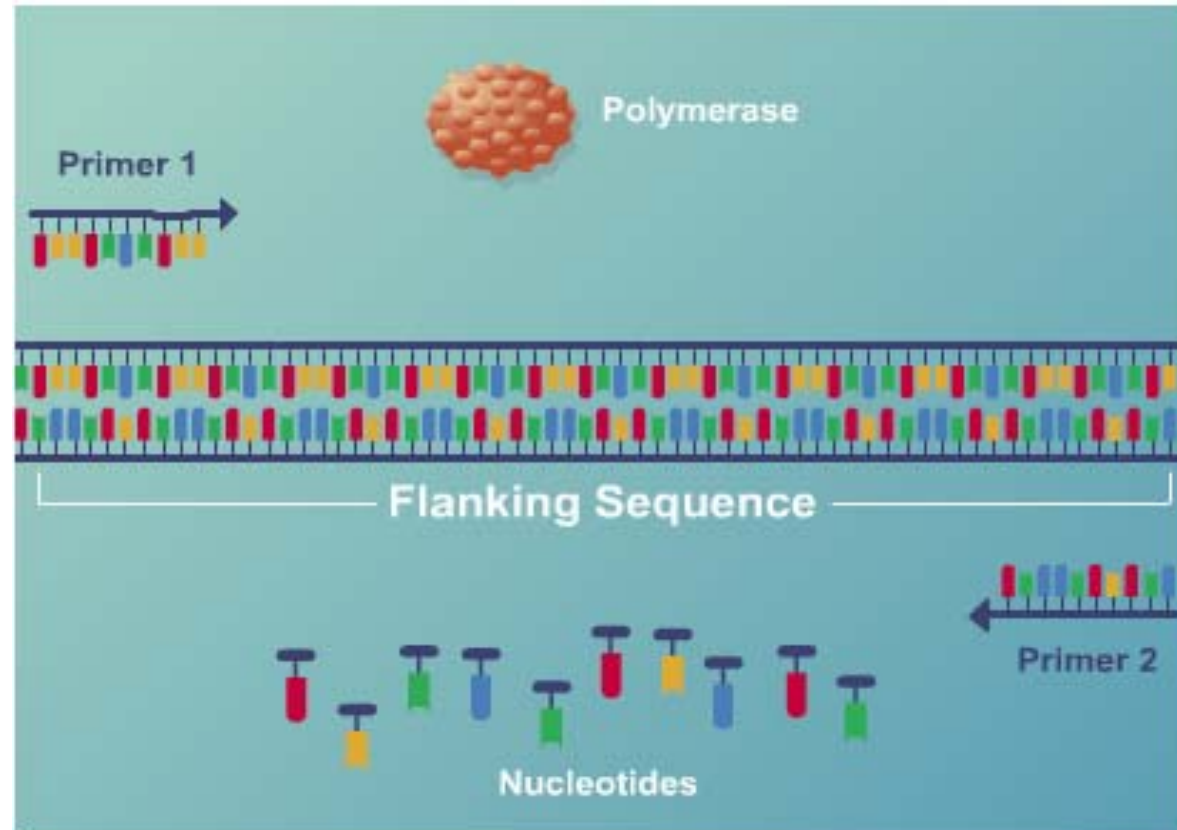
- 0005F
- 1540R
- 12 Seq Primers

500bp

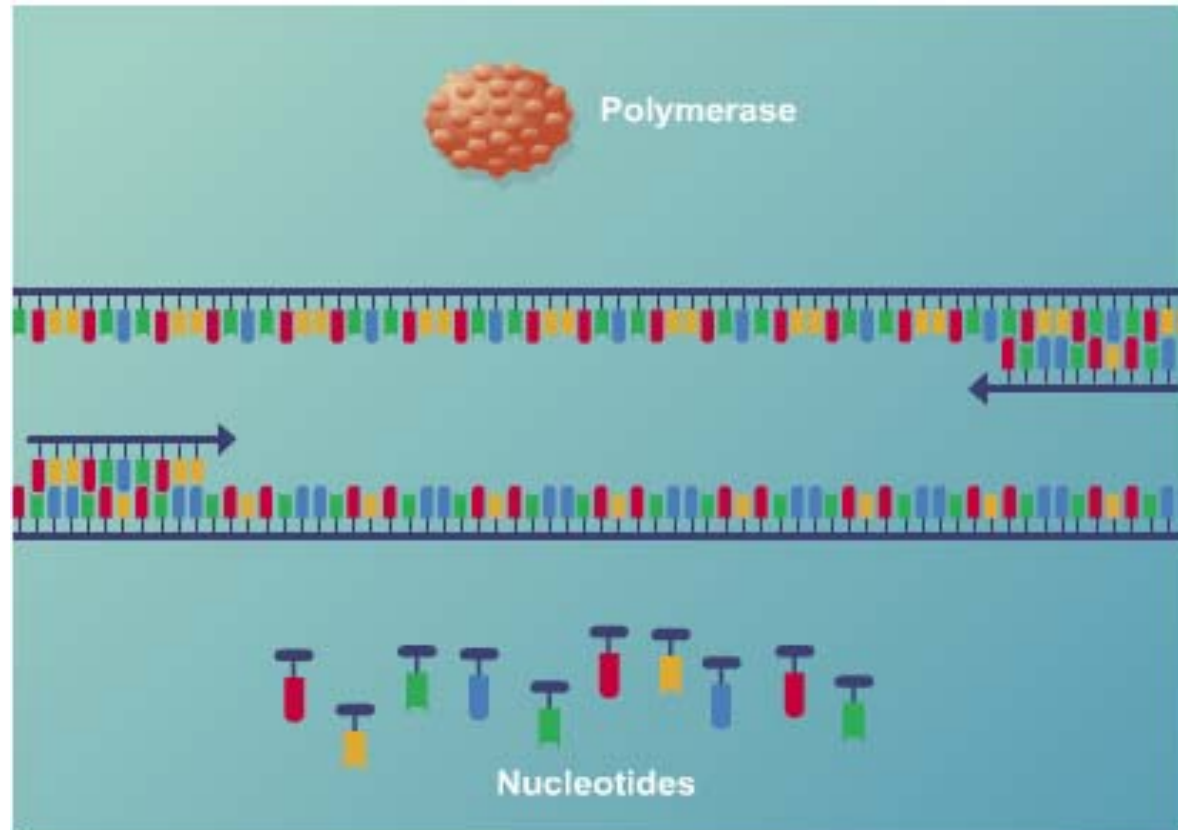
- 0005F
- 0531R
- 2 Seq Primers



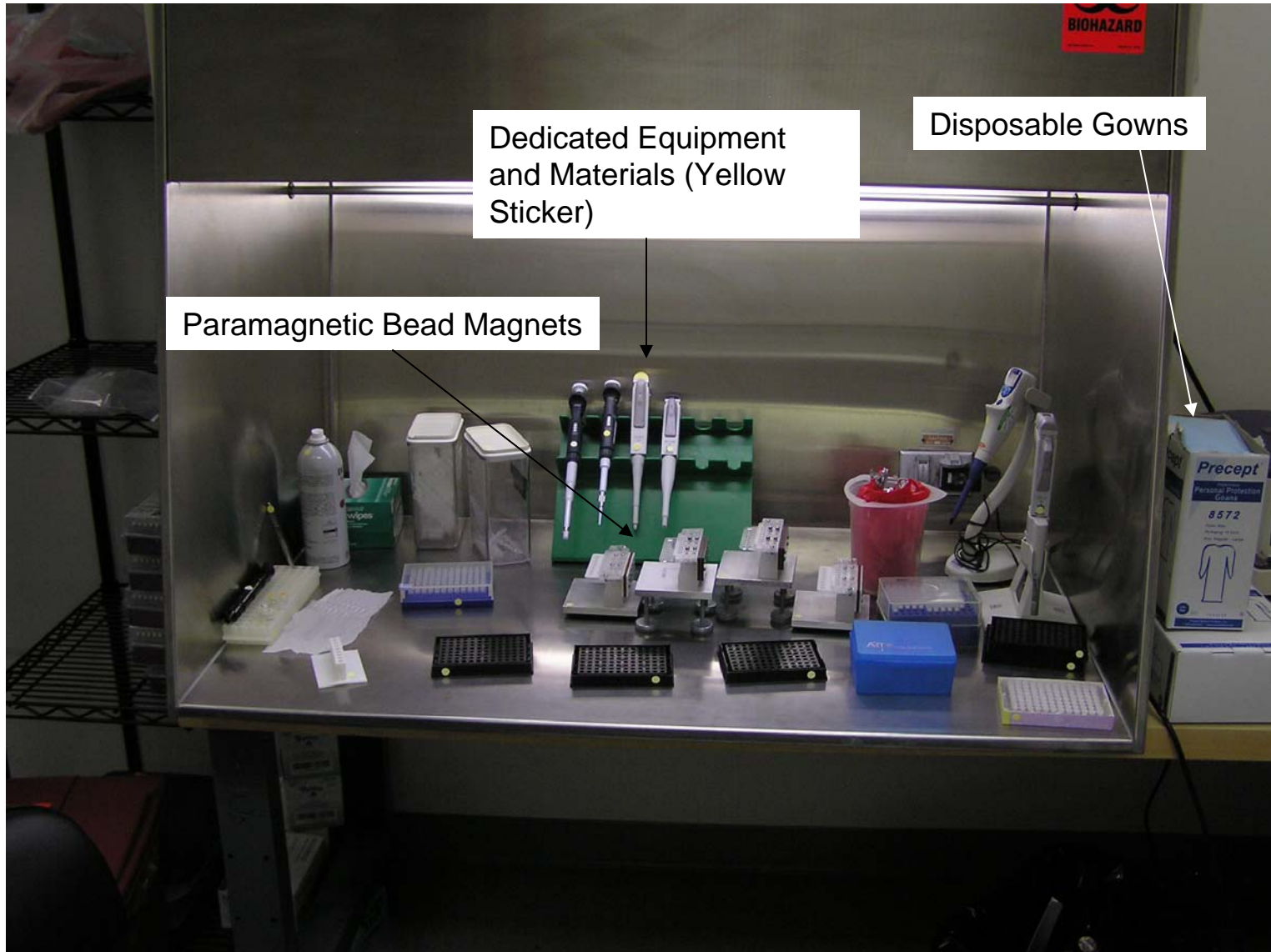
PCR



PCR



PCR



Dedicated Equipment
and Materials (Yellow
Sticker)

Disposable Gowns

Paramagnetic Bead Magnets



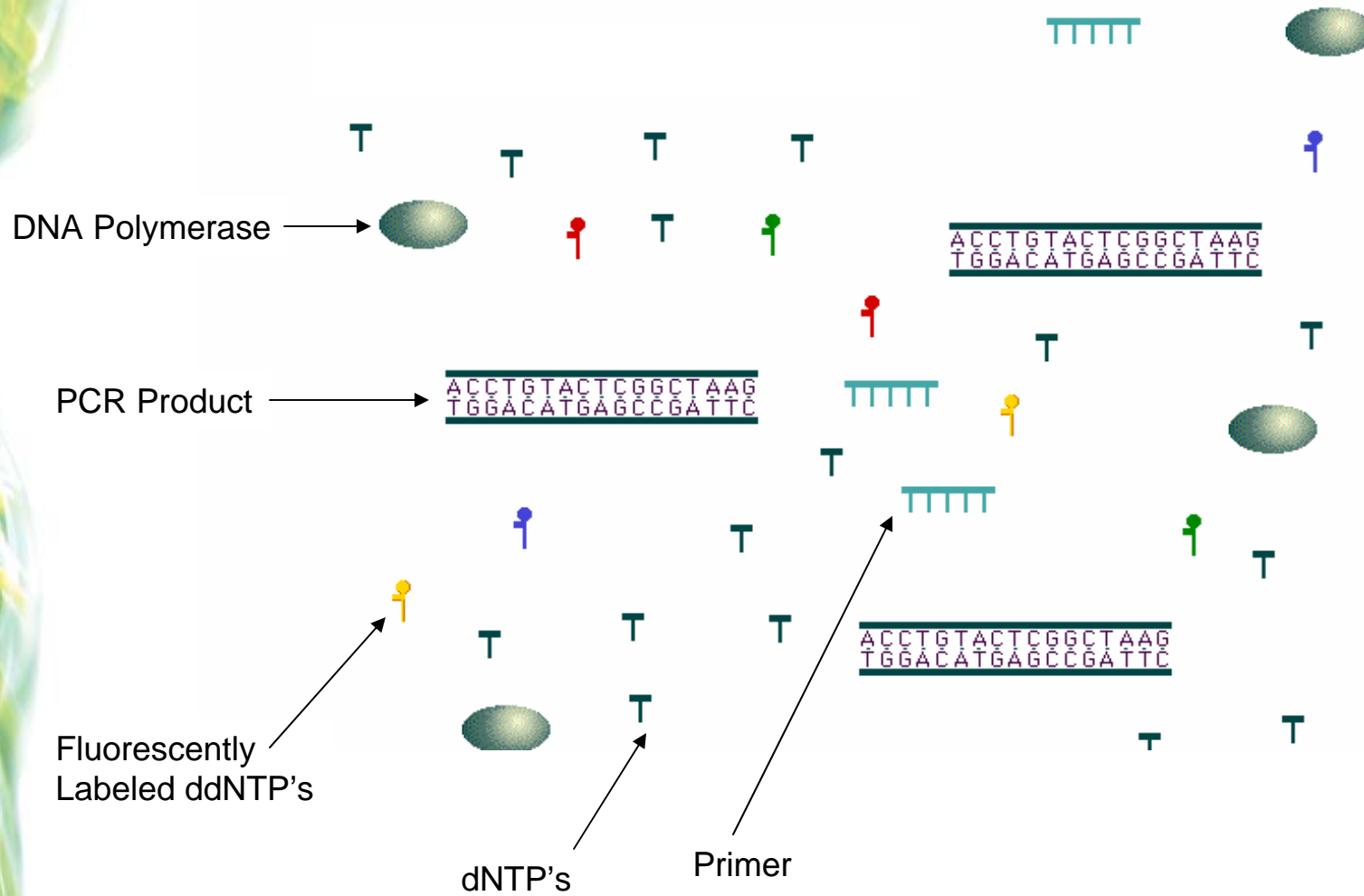
PCR Clean-up

- Exo-SAP
 - Enzymatic clean-up step
 - Exonuclease removes excess primers from PCR step
 - Shrimp Alkaline Phosphatase (SAP) removes terminal phosphate from excess dNTP's
- Exo-SAP clean-up results in less amplicon loss than filtration or size exclusion steps

Cycle Sequencing

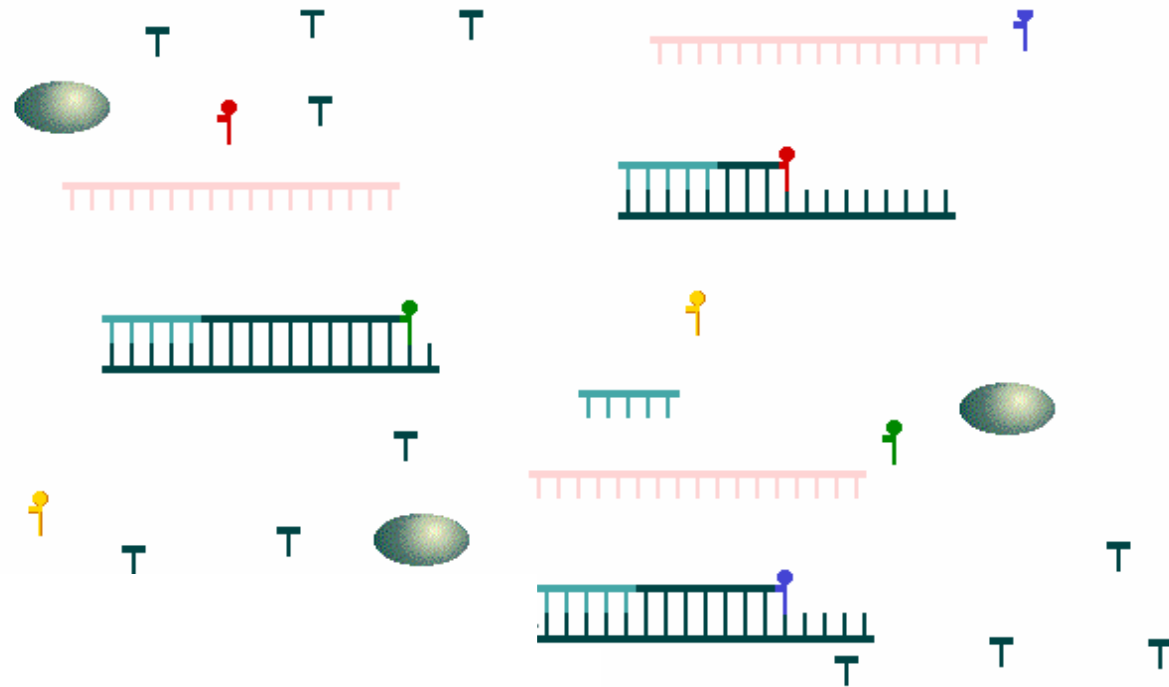
- Bacterial and Fungal Master Mixes using latest ABI fluorescent dye chemistry
 - DNA Polymerase
 - dNTP's
 - Fluorescently labeled, ddNTP's
 - 1 Primer per reaction
- Purified PCR product
- Generation of extension products, which are used to determine DNA sequence of PCR product

Cycle Sequencing





Cycle Sequencing

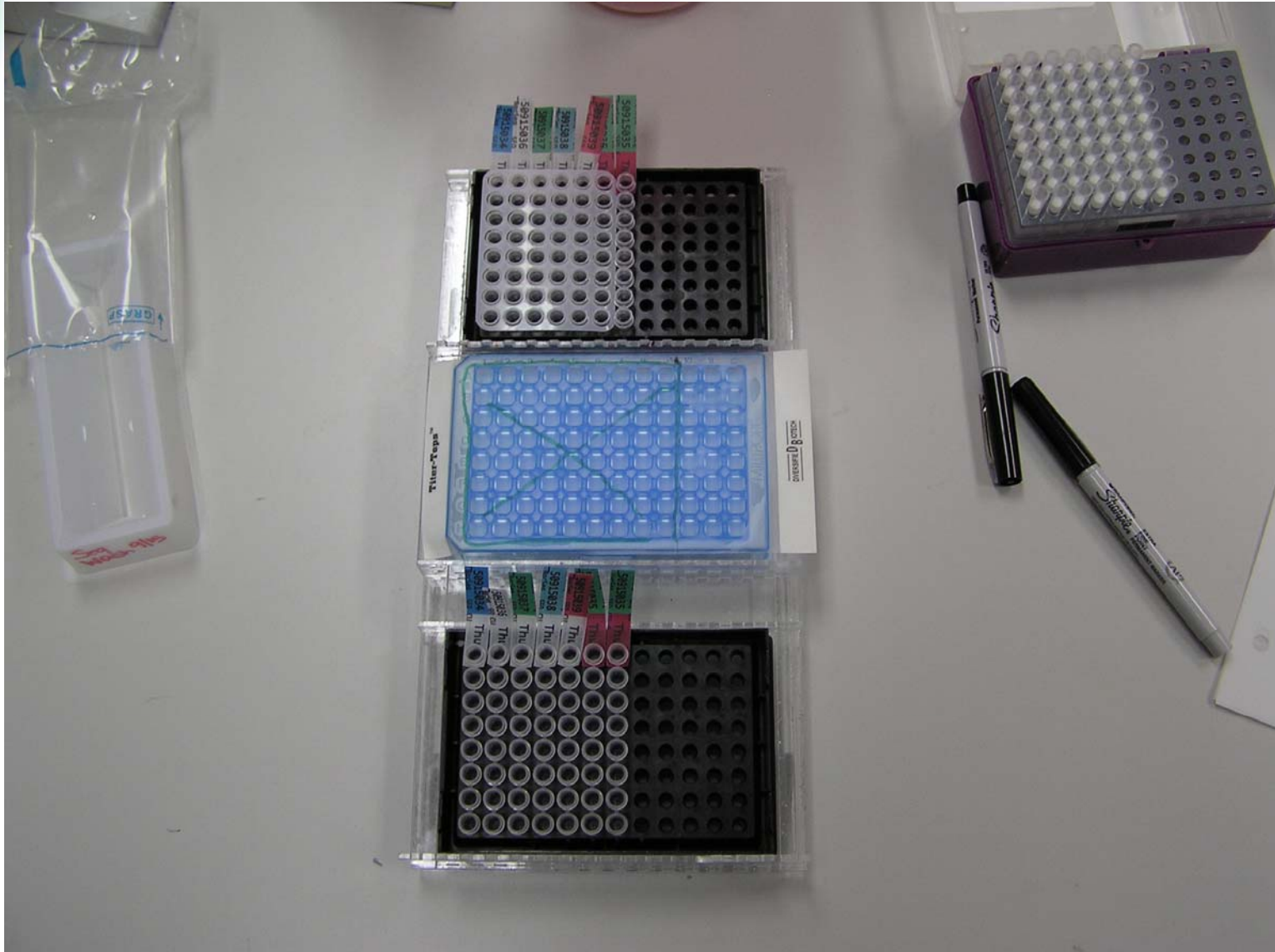
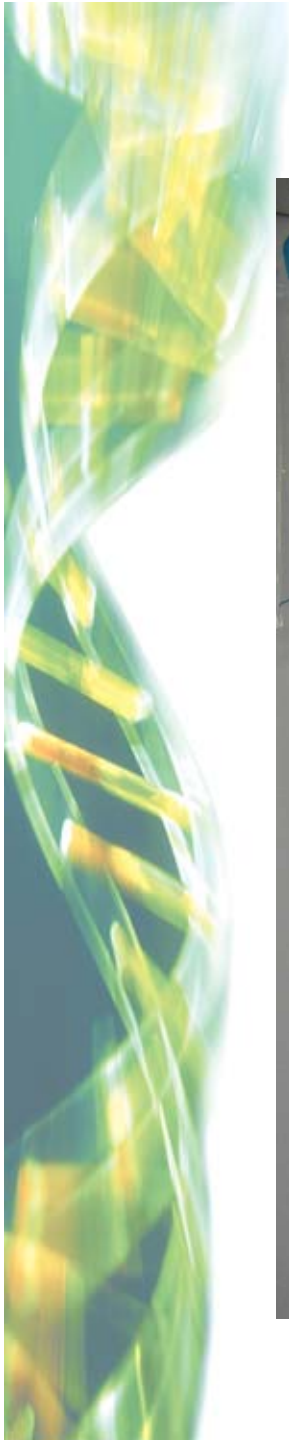


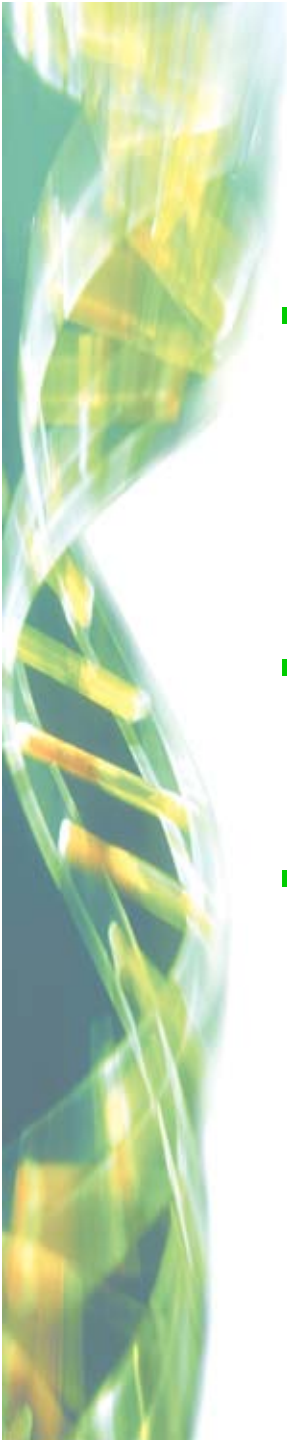


Cycle Sequencing Clean-up

- Extension products are purified using size based filters
- Filters allow unincorporated primers, dNTP's, and ddNTP's to pass through
- Extension products remain on the filter
- Extension products are washed with water, then resuspended in Injection Solution (low conductivity)

Cycle Sequencing Clean-up

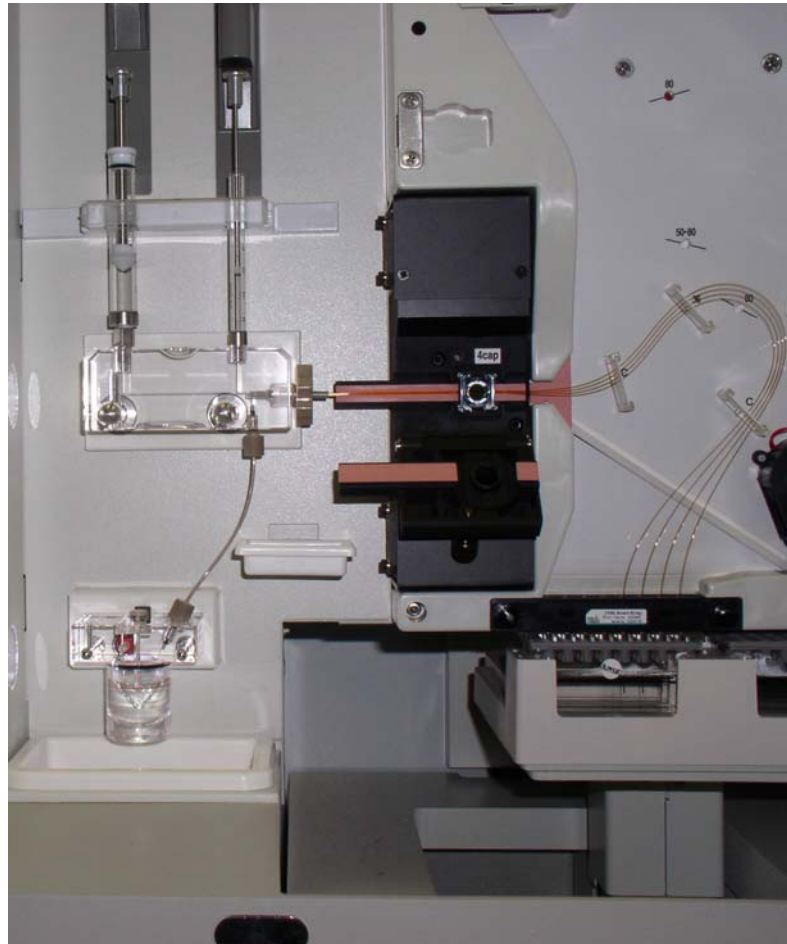




Automated Fluorescent DNA Sequencing

- Size separation of extension products from Cycle Sequencing reaction
 - ABI 3100 - 16 channels
 - Run time of 1 hour
- Sequencer automatically injects, separates, and detects extension products
- Generates Electropherogram of Sequence data

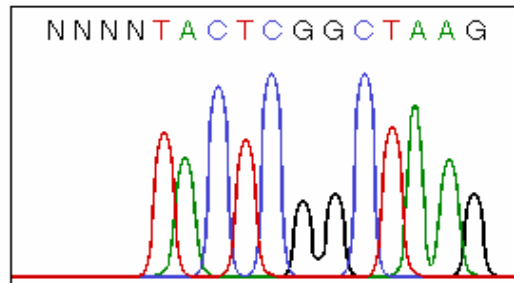
ABI 3100 Genetic Analyzer



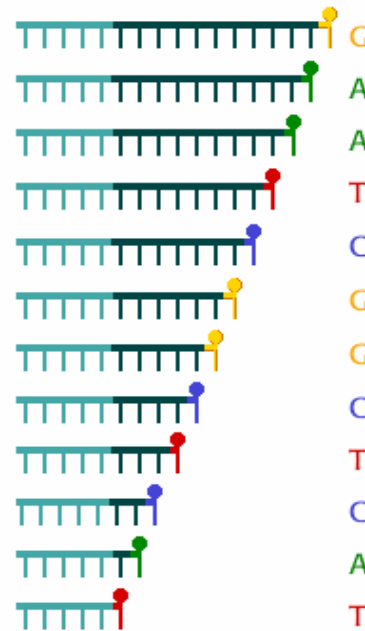
Automated Fluorescent DNA Sequencing

Cycle Sequencing

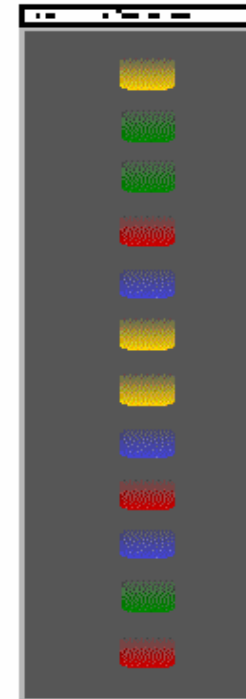
The simulated gel image is read from bottom to top, starting with the smallest fragment.



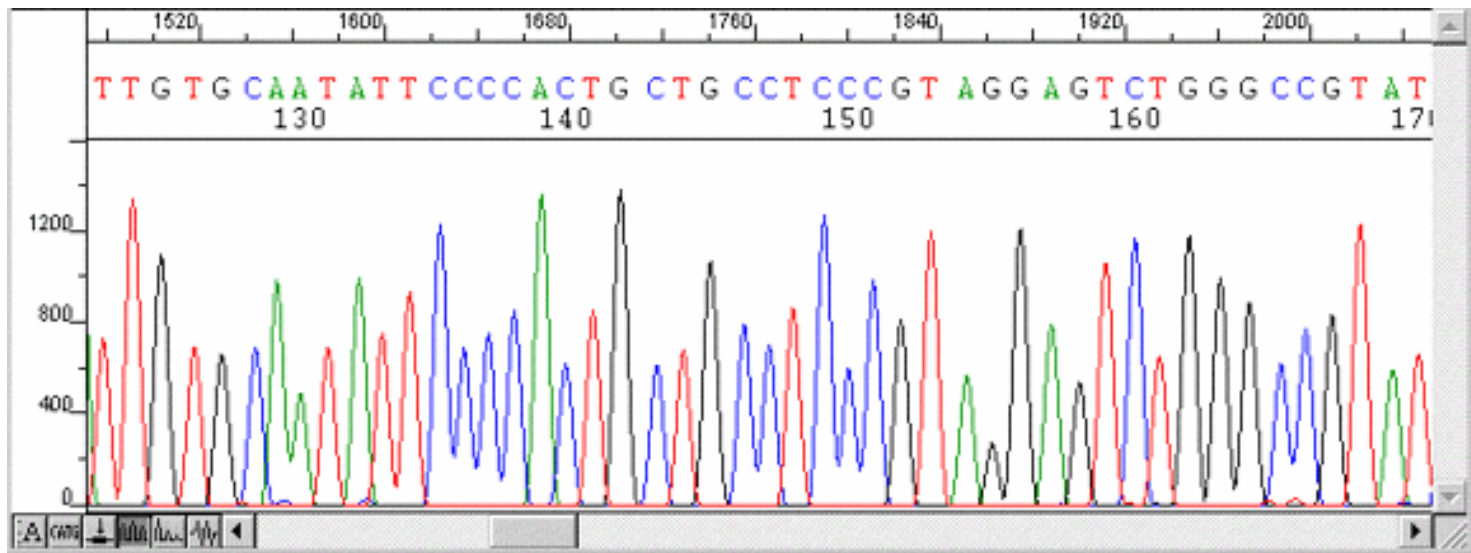
GO TO START



QUIT



Automated Fluorescent DNA Sequencing Analyzed Data (Electropherogram)





Identification Report - Bacterial 500bp

Customer: Example Report

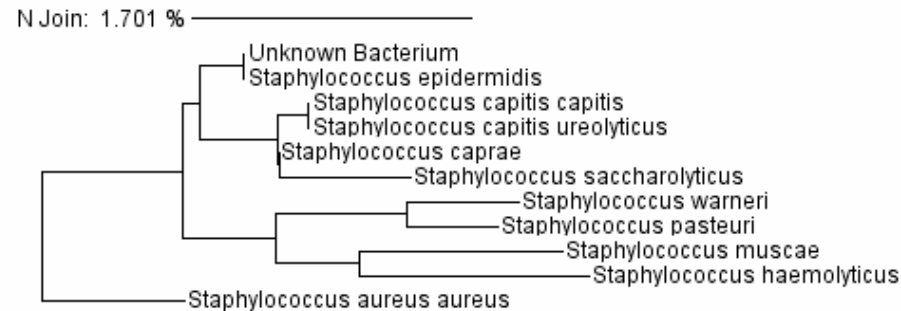
Sample: Unknown Bacterium

Date: Friday, November 8, 2002 11:34 AM



Alignment: 535 Unknown Bacterium
0.00 % 535 Staphylococcus epidermidis
0.75 % 535 Staphylococcus caprae
0.93 % 535 Staphylococcus capitis ureolyticus
0.93 % 535 Staphylococcus capitis capitis
1.54 % 535 Staphylococcus saccharolyticus
2.24 % 535 Staphylococcus warneri
2.24 % 535 Staphylococcus aureus aureus
2.24 % 535 Staphylococcus muscae
2.43 % 535 Staphylococcus pasteuri
2.62 % 535 Staphylococcus haemolyticus

Neighbor Joining Tree



MicroSeq™ Database Search Result

Identification: Staphylococcus epidermidis

Confidence Level: Species

Reviewed _____

