



TR40 Sterilizing Filtration of Gases A comparison with TR26 Sterilizing Filtration of Liquids

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Sterilizing Filtration of Gases

- Published Jan/Feb 2005
- Educational guide to complement TR26
- Committee
 - F. Bing (Chair), S. Sundaram (Co-chair)
 - B. Bardo, T. Britton, R. Conway, T. Feeser,
 - H. Haughney, A. M. Jones, M. Jornitz, S. Langille
 - R. Levy, R. Madsen, J. Martin, L. McBurnie
 - T. Meltzer, D. Meyer, G. Morris, D. Ridealgh
 - H. Schroeder, P. Stinavage, A. M. Trotter
 - 7 manufacturers, 6 users, 4 consultants, 1 FDA

The background of the slide is a light blue-tinted photograph of various laboratory glassware, including beakers, flasks, and bottles, arranged on a surface. The items are slightly out of focus, creating a professional and scientific atmosphere.

Similarities

- Both technical reports are considered to be educational guides rather than mandatory or implied standards
- Both describe filter retention mechanisms, selection criteria, sterilization methods, validation of retention capabilities and integrity test methods

Differences

- The risk associated with liquid filtration is significantly greater than the risk associated with gas filtration
 - Bioburden potential is higher in liquid
 - Example suggested action levels
 - 100 cfu mL for Purified water (liquid)
 - 100 cfu/M³ for class 8 /100,000 cleanroom
= 0.0001 cfu/mL (air)

Removal mechanisms

- Gas filters have additional retention mechanisms and will retain smaller particles

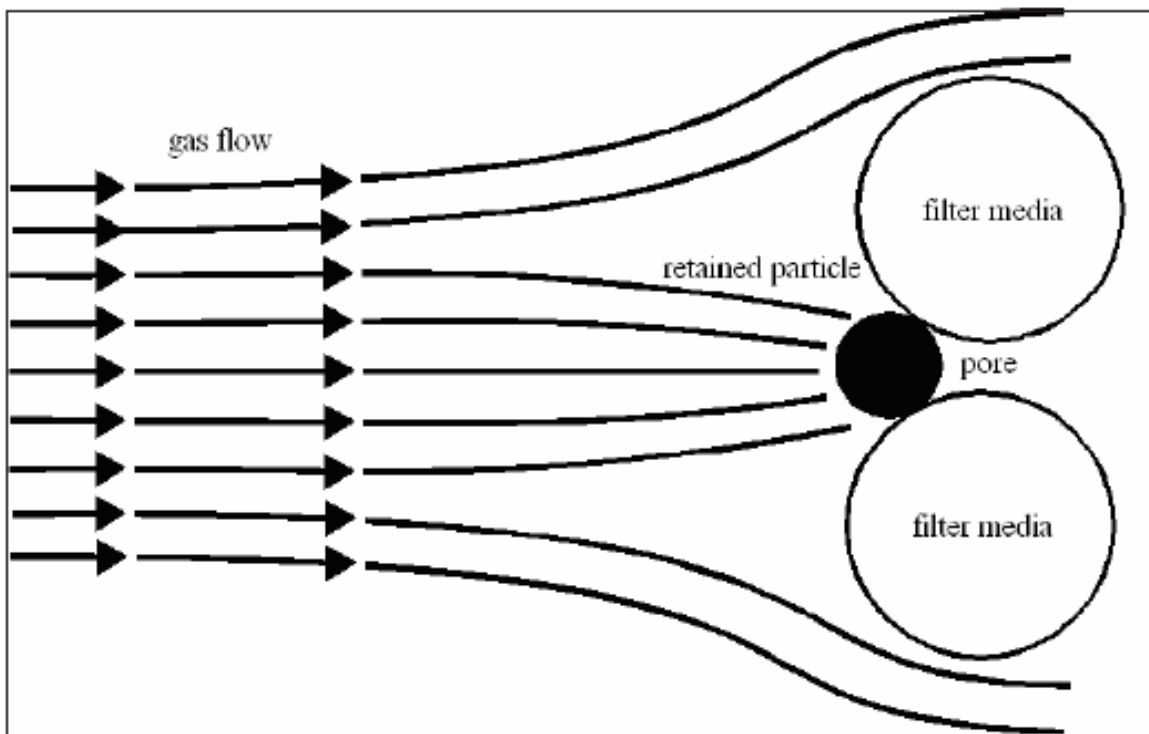


Figure 1: Particle Retention by Size Exclusion

–Size exclusion is used in both liquid and gas

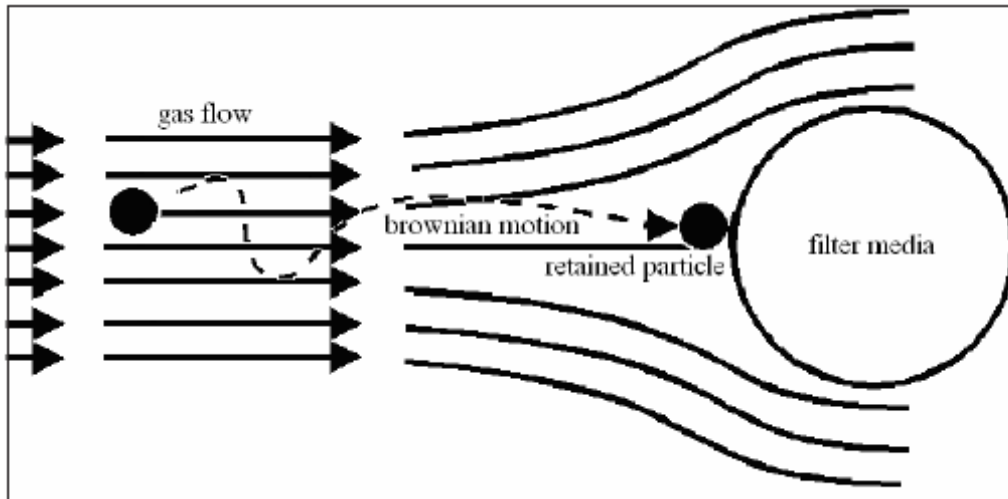


Figure 2: Diffusional Interception

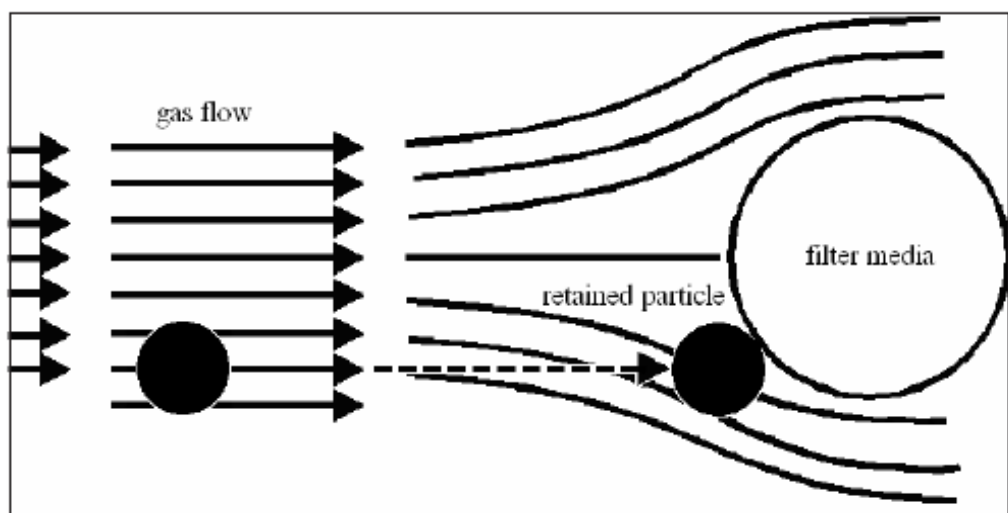


Figure 3: Inertial Impaction

- Smaller particles in gas
 - Diffusional interception
 - Electrostatic attraction
 - Inertial impaction

Most penetrating particle size

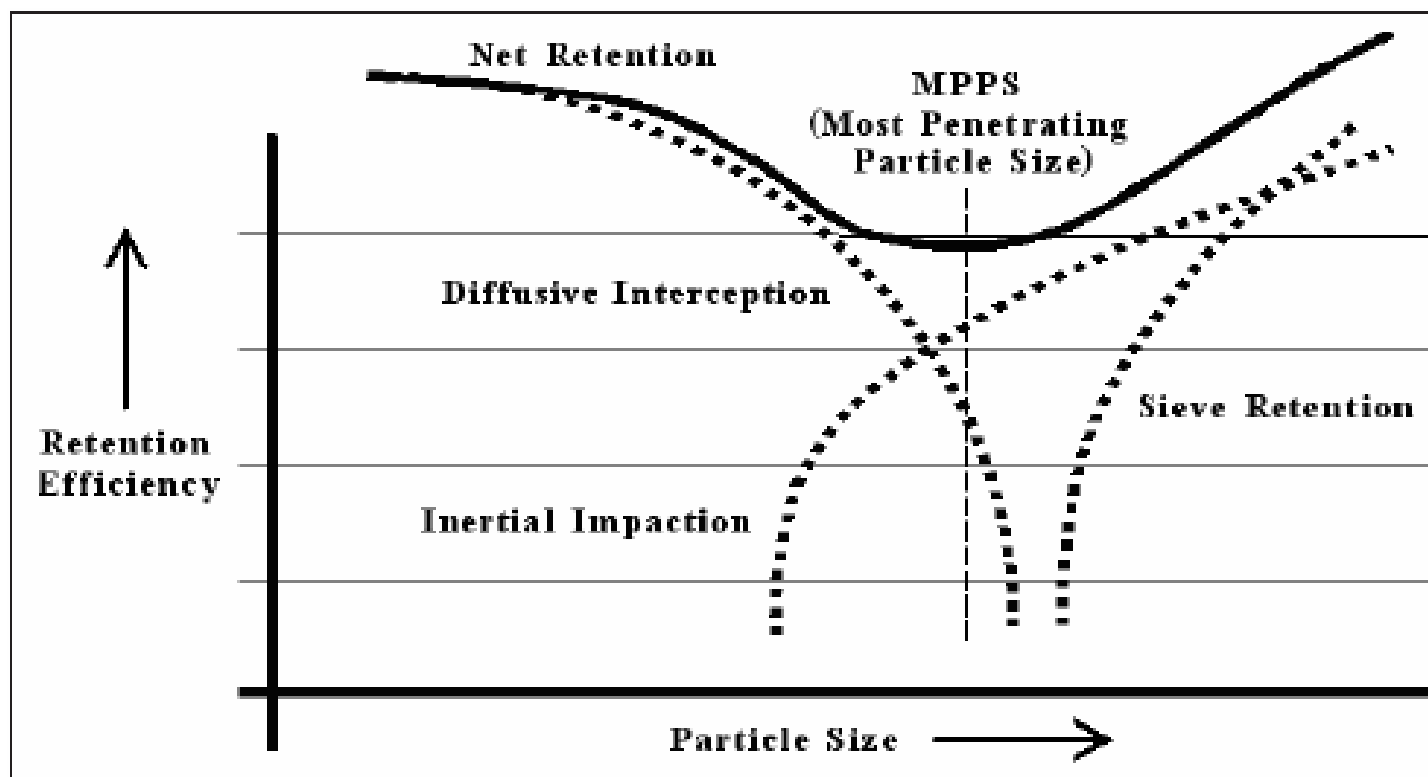


Figure 4: Effect of Various Retention Mechanisms of Particles Retained from a Gas Stream as a Function of Particle Size

Hydrophobic membranes

- Do not readily wet with water and so avoid water blockage that can occur with hydrophilic membranes

Polymer	Critical surface tension (dynes/cm)
PTFE	18
PVDF	25
Polypropylene	29.5
Polyethylene	32

Pore size ratings

- True pore size should not be confused with nominal pore rating given by the manufacturer
- Even less meaning in the ratings of gas than liquid filters
- Gas filters are best described by performance on a challenge test correlated to a filter integrity test
- Liquid rated “sterilizing grade” 0.2 μm are much more efficient in retention in dry gas streams

Filter selection criteria

- Retention capacity
- Integrity testing
- Flow rate & throughput
- Materials of construction
 - Hydrophobicity
 - Durability
 - Toxicity
 - Particle shedding
 - Compatibility

Design considerations

- Minimize water blockage
 - Orient housing to allow condensation to drain
 - Jacket or heat trace housing (3-5°C above process temperature)
 - Open vent valve
 - Coalescing prefilter
- Integrity test
 - Test considerations
 - Wetting
 - Drying (blow down)

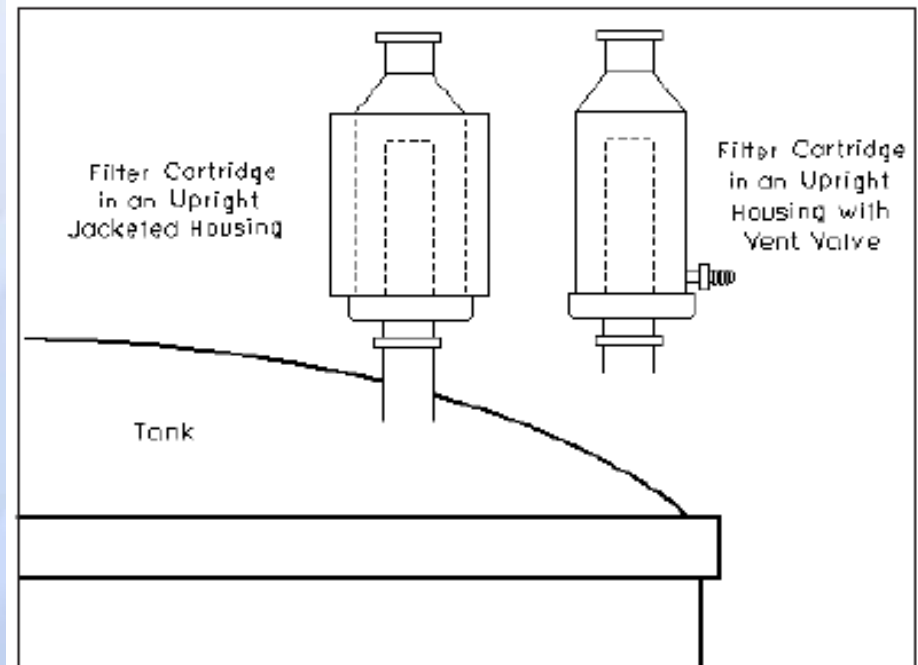


Figure 6: Examples with Filter Oriented for Drainage of Condensate with Steam Jacket or Open Lower Vent Valve

Ideal sterile gas filter

- Retains microorganisms even under adverse conditions such as high humidity
- High thermal /mechanical resistance
- Withstand multiple steam cycles
- High gas flow at low ΔP
- Hydrophobic
- Non fiber releasing
- Integrity testable – correlated to removal efficiency
- Easy to install and maintain
- Compatible with application

Most critical applications

- Gas is in contact with sterile final product or critical surfaces of the associated equipment
 - Compressed process gases for aseptic fill operations
 - Vacuum break gases for lyophilizers and critical autoclaves
 - Headspace gases used to flush vials and ampoules
 - Sterile bulk holding tank vents
 - Nitrogen blankets
- Filter should be qualified with a liquid based bacterial challenge test and have a physical integrity test correlated to retention in liquid

Moderately critical applications

- Filtered gas is not in direct contact with exposed sterile product or surfaces
 - Intermediate process steps
 - Air supplied to a fermentation process
- Filters qualified with aerosol based bacterial challenge, correlated to a physical integrity test, are appropriate

Other applications

- Applications that only require a reduction in bioburden have less stringent requirements
- Because the retention expectation is similar to HEPA filters, dispersed oil aerosol challenges are deemed acceptable to establish the retention capability



Special cases

- Some applications may have additional or more specific requirements
 - e.g. bacteriophage control or virus retention
- Different articles have been published regarding retention of contaminants bacteria, phages under different conditions
- Applicability of the data to the particular situation needs to be evaluated on a case by case basis

Validation of retention capabilities

- No specific standard that defines the retention requirements of a membrane filter used to sterilize gases
- Several approaches
 - Liquid challenge
 - Aerosol challenge
 - bacteria, spores, virus, dispersed oil
- Retention studies do not need to be repeated by user
- Should evaluate applicability of the retention study to the application

Liquid challenge

- Liquid bacterial challenge represents the worst-case condition since retention in liquids is lower than gases
- ASTM F838 or comparable test on discs, capsule or high area cartridge
 - *Brevundimonas diminuta* ATCC® 19146™
 - 100% of effluent must be analyzed

Aerosol challenges

- Bacterial (spore) aerosol challenges are always less rigorous than liquid challenges even though they do represent the way the filter is challenged in a dry gas process
- Phage/viral challenges may be the least rigorous because in gas filtration, the smallest particles are not the most difficult to retain

Aerosol bacterial challenge

- *B. diminuta* - watch conditions for viability
- *Bacillus subtilis* - spores resist drying but are larger than vegetative cells
- Nebulizer generates droplets
- Andersen Sampler can be used to assess droplet size down to 0.65 μm
- Filtered gas is analyzed using liquid impingers
- Control, without filter is run to determine the challenge level
- Lower flow rates may be worst-case scenario

Sample aerosol challenge apparatus

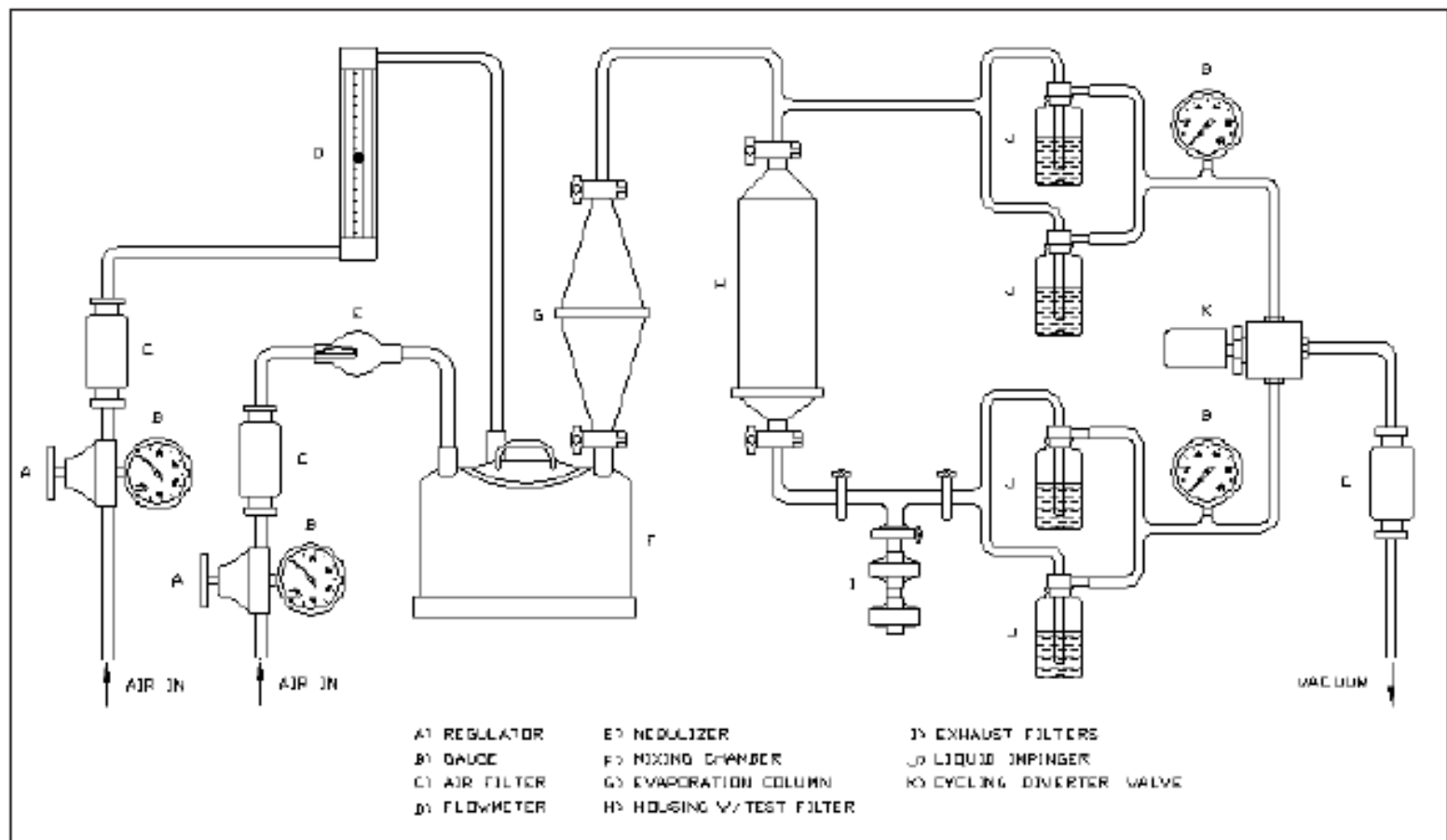


Figure 7: Schematic of Aerosol Challenge Test (Split Stream Approach)

Viral aerosol challenges

- No standards
- Similar apparatus
- Bacteriophage: Phi X-174, PP7, MS2, T1
- Virus sizes = 25 nm to 180 nm
- Andersen Sampler can demonstrate droplets are <650 nm
- MPPS tends to be in 200-300 nm range
- Viral aerosol challenges may be least rigorous microbial challenge

Viral aerosol challenge

- Challenge size may be larger than virus depending on drying and size cannot be precisely established
- Viral spike solution is typically prefiltered to remove aggregates (0.2-0.1 μm)
- Higher flow rates may be worst-case since they diminish diffusional interception
- Impinger fluid is analyzed for the test particle with an infectivity assay
- A presence/absence test can be performed on the remaining fluid

Integrity tests

- Retention challenges should be correlated to an integrity test
- Traditional wetted membrane tests using a low surface tension fluid
 - Bubble Point Test
 - Diffusive/Forward Flow Test
 - Pressure Hold/Decay Test
- Water Intrusion Test (WIT)
- Aerosol Integrity Test



WIT

- Water Intrusion Test does not involve wetting the membrane with solvent
- The upstream side of the filter is flooded with water, pressurized and allowed to stabilize ~10 minutes
- The flow of water vapor through the membrane is measured over time
- Useful test for new filters, filter must be dry prior to testing

Aerosol integrity test

- Historically used for detecting failures in HEPA and ULPA grade filters
- Filter is challenged with $10^7/\text{cm}^2$ 0.2-0.3 μm aerosol generated from highly refined mineral oil
- A downstream sensor (laser particle counter) detects oil droplets that penetrate the filter
- Can be correlated to aerosol microbial challenge

When to integrity test

- Before sterilization – right filter, correctly installed
- Post sterilization – also detects if the filter was damaged during sterilization
- Post use – confirms filter remained good throughout the critical process

Extended use applications

- Parallel filters, use one while other is being tested and prepared for use
- Redundant filters with periodic testing and change-out
- Combination of periodic testing and change-out
- Test once only, after the first sterilization
- Do not test filters and base change-out on historical data (# sterilization cycles or time on line)

User validation of critical applications

- Generic data correlating retention (bacterial or viral) to the integrity test
- Qualification data for toxicity, durability, compatibility, recommendations for integrity test parameters
- Evaluate retention data applicability to process
 - liquid-rated represents worst case
- Physical integrity test
- Compatibility and service life in use
 - May be demonstrated by integrity testing filter

Criteria	Filter user	Filter manufacturer
	Device	Disc /Device
Bacterial retention/integrity test relationship data	(E)	(Q)
Integrity test	-	(Q/R/L)
Integrity test method & selection	(E)	(R)
Microbial/viral retention (liquid/aerosol)	(E)	(Q/L)
Compatibility/ service life	(E/V)	(Q/R)
Toxicity testing	-	(Q)
Effects of sterilization method on filter integrity	(E/V)	(Q)